

(11) **EP 0 780 475 B1**(12) **EUROPEAN PATENT SPECIFICATION**

(45) Date of publication and mention
of the grant of the patent:
09.06.1999 Bulletin 1999/23

(21) Application number: 95112559.0

(22) Date of filing: 09.08.1995

(51) Int Cl.⁵: C12N 15/86, C12N 15/45,
C07K 14/12, C12N 7/00,
C12Q 1/68, C12N 15/62,
C07K 14/145, A61K 48/00
// C12N9/00, A61K39/165

(54) **CDNA corresponding to the genome of negative-strand RNA viruses, and process for the production of infectious negative-strand RNA viruses**

Dem Genom von minussträngigen RNA-Viren entsprechende CDNA und Verfahren zur Herstellung von infektiösen minussträngigen RNA-Viren

ADNC correspondant au génome des virus à ARN brin-négatifs et procédé pour la production des virus à ARN brin-négatifs infectieux

(84) Designated Contracting States:
AT CH DE DK FR GB IT LI NL SE

(43) Date of publication of application:
25.06.1997 Bulletin 1997/26

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Remarks:

The file contains technical information submitted after the application was filed and not included in this specification

Description

BACKGROUND OF THE INVENTION

Technical Field

[0001] The present invention relates, in general, to a methodology for the generation of nonsegmented negative-strand RNA viruses (Pringle, 1991) from cloned deoxyribonucleic acid (cDNA). Such rescued viruses are suitable for use as vaccines, or alternatively, as vectors in somatic gene therapy applications. The invention also contemplates for its practise cDNA molecules suitable as tools in this methodology and helper cell lines allowing the direct rescue of such viruses. Measles virus (MV) is used as a model for other representatives of the *Mononegavirales*, in particular the family *Paramyxoviridae*.

[0002] The invention provides the technology for construction of recombinant vaccine strains, in particular MV vaccine strains containing coding regions for the expression of epitopes or entire protein from other viruses, bacteria, or parasites. It also demonstrates that chimeric MV strains containing heterologous envelope proteins can be constructed suitable for targeting cells not containing an MV receptor. Thus, in principle, plasmids based on the genome of MV, packaged in envelopes containing proteins for targeting special cell types can be constructed, encoding gene products either lacking in genetically defective individuals or toxic for targeted malignant cells.

[0003] By straightforward replacement of the MV-specific helper cell lines described in this invention by cell lines expressing the cognate proteins encoded by other representatives of the *Mononegavirales* to be rescued, any other member of this viral order replicating in vertebrate cells can be used for the purpose of live vaccines or of vectors for gene therapy instead of MV.

Background Information

Measles virus

[0004] MV is a member of the family *Paramyxoviridae*. Its genetic information is encoded on a single RNA strand of negative polarity, comprising 15894 nucleotides. The genome is sequentially transcribed from the 3' terminus to yield, in addition to a leader RNA, 6 major capped and polyadenylated messenger ribonucleic acid (RNA) species, each of which encodes one major protein. The genome map is shown in Figure 1, indicating the genes specifying as the principal products N (nucleocapsid protein), P (phosphoprotein), M (matrix protein), F (fusion protein), H (hemagglutinin) and L (large protein = polymerase). Several additional RNA and protein species, in part mentioned in the Table of Fig. 1 complicate this simple picture, but they are not relevant here.

[0005] MV is a major cause of acute febrile illness in

infants and young children. According to estimates of the World Health Organisation (WHO), one million young children die every year from measles. This high toll arises primarily in developing countries, but in recent years also industrialised countries such as the USA have been affected again by measles epidemics, primarily due to incomplete adherence to immunisation programs (Clements and Cuttis, 1995). At present, several live attenuated MV vaccine strains are in use (including the Schwarz, Moraten and Edmonston-Zagreb strains), almost all derived from the original Edmonston strain (Enders and Peebles, 1954) by multiple passage in non human cells (Enders, 1962). For a recent discussion of MV vaccinology including future trends see Norrby (1995). Measles vaccine is usually administered at 15 months of age or, in developing countries, already at 6 months, and has proved to be highly effective, usually providing life-long immunity against MV reinfection eliciting morbidity. To date, the genetic alterations responsible for attenuation of these vaccine strains remain unknown. The proven safety of measles vaccine, combined with its high and long-lasting efficiency, predetermines it as an ideal plasmid for the expression of heterologous genes. Such a vaccine may prove as efficient in eliciting long-lasting immune protection against other pathogenic agents as against the vector virus itself. Another possible candidate as vaccination vector is Mumps virus, a distant relative of MV, which is also highly efficacious and safe as attenuated live vaccine.

Rescue of RNA virus from cloned DNA

[0006] The study of the replication cycle of a number of RNA viruses has been greatly facilitated by the availability of DNA clones from which infectious virus can be rescued, thus allowing the application of reverse genetics. Initially, the bacteriophage Q β (Taniguchi et al., 1978) and polio virus (Racaniello and Baltimore, 1981), and subsequently Sindbis virus (Rice et al., 1987) were expressed from cloned cDNA. To date, a large variety of positive-strand RNA viruses, primarily infecting vertebrates and plants, can be rescued from cloned DNA (for a recent review see Boyer and Haenni, 1994). In addition, proviral DNA of retroviruses is infectious. However, attempts to obtain infectious virus from cDNA clones of negative-strand RNA viruses have met with great difficulties. This is due to two properties of these viruses: (i) neither genomic nor antigenomic RNAs are infectious, because they do not serve as mRNAs; and (ii) both transcription and replication require ribonucleocapsids, i.e., rod-like nucleoprotein complexes (RNPs), containing the genomic RNA and several proteins with structural and/or enzymatic function.

[0007] Rescue from cloned DNA has been achieved several years ago in the case of influenza virus, a negative-strand RNA virus containing eight genome segments. Their RNPs which are small in size and loosely structured as revealed by the susceptibility of their RNA

component to RNase, can be assembled in vitro from RNA and the required viral proteins, N and the polymerase components. Initially, an artificial RNA has been used carrying as a reporter the chloramphenicol acetyltransferase (CAT) coding sequence embedded in the noncoding terminal segments of an influenza virus genome subunit (Luytjes et al., 1989). Later, single authentic or altered genome subunit RNAs transcribed in vitro from cloned DNA were used (Enami and Palese, 1991). The assembled RNPs replicated and transcribed upon transfection into influenza-infected cells, as monitored by CAT production and by rescue of a reassorted influenza virus, respectively. Purification of virus containing the introduced subunit from the vast excess of non-reassorted virus in some cases can be accomplished by selection, for example, using a specific neutralising antibody directed against the protein encoded by the cognate subunit of the helper virus.

[0008] In contrast, for the viruses with a nonsegmented negative-strand RNA genome, grouped together in the order *Mononegavirales* (Pringle, 1991) the much more tightly structured and longer RNPs, containing in addition to the N protein the assembly and polymerase cofactor phosphoprotein (P) and the viral RNA polymerase (large protein, L) have been refractory to functional reassortment in vitro. Therefore, many laboratories approached the rescue of representatives of the *Mononegavirales* starting out with subgenomic RNAs containing only essential sections of the viral genomes, using viruses to provide the helper proteins required to intracellularly encapsidate and replicate these mini-replicons. First, naturally arising subgenomic RNAs, competing with the viral replication and thus known as defective interfering particle (DI) RNAs (Re, 1991) were used, being substituted later by artificial DI RNAs containing reporter genes, transcribed from appropriately constructed plasmids. These mini-replicons, first devised by the group of M. Krystal (Park et al., 1991) according to the replicon used for the initial influenza rescue model (Luytjes et al., 1989), carry a CAT coding sequence inserted into viral noncoding terminal regions of Sendai virus (SeV) and have been used successfully also for respiratory syncytial virus (Collins et al., 1993; Collins et al., 1991), human parainfluenza virus 3 (Dimmock and Collins, 1993), rabies virus (RV) (Conzelmann and Schnell, 1994) and MV (Sidhu et al., 1995). [0009]. In all these systems, the essential helper proteins were provided either by the homologous viruses or by the vaccinia vector vTF7-3 encoding phage T7 RNA polymerase (Fuerst et al., 1986) to drive T7-specific transcription of transfected plasmids encoding the required proteins N, P and L as pioneered by Pattnaik et al., (1990). These investigations using mini-replicons have allowed important insights into the noncoding regulatory regions of the corresponding viral genomes and antigenomes (for a recent discussion see Wertz et al., 1994). Adopting the same experimental set up, the rescue of VSV, as RV a member of the *Rhabdoviridae*, has

now also been reported (Lawson et al., 1995).

[0010] An important drawback of that method (as well as the method reported for the rescue of negative-strand RNA viruses with a segmented genome) is the involvement of a helper virus which has to be separated from the rescued virus and which can interfere with the replication of the virus to be rescued. For RV and VSV, both belonging to the rigidly structured *Rhabdoviridae* and replicating to high titers, this is not an important problem. However, in case of loosely structured, polymorphic virions typical for the members of the family *Paramyxoviridae* and in case of viruses yielding only relatively low titers, the presence of a helper virus would render the recovery of rescued viruses difficult and may well preclude their rescue altogether.

[0011] Accordingly, the technical problem underlying the present invention was to provide genetic material useful for the generation of non-segmented negative-strand RNA viruses, preferably of the family *Paramyxoviridae* and most preferably of measles virus and a system for the recovery of such viruses with reasonable efficiency. The solution to said technical problem is provided by the embodiments characterised in the claims.

[0012] Thus, the present invention relates to a method for the production of an infectious non-segmented negative-strand RNA virus of the order *Mononegavirales* comprising

- (a) introducing a cDNA molecule comprising the entire (+)-strand sequence of said negative-strand RNA virus operatively linked to an expression control sequence, which allows the synthesis of antigenomic RNA transcripts bearing the authentic 3'-termini, into a helper cell expressing an RNA-polymerase, preferably T7 RNA-polymerase, an N and a P protein, preferably of the virus to be rescued, and, further, an L protein, preferably of the virus to be rescued, encoded by a cDNA comprised by a plasmid either transiently or stably introduced into said cell; and
- (b) recovering the assembled infectious non-segmented negative-strand RNA virus.

[0013] Accordingly, the present invention can be practised by the provision of a cDNA molecule for the production of any negative-strand RNA virus of the family *Paramyxoviridae*. Preferably said antigenomic RNA transcripts also bear the authentic 5'-termini.

[0014] As has been further found in accordance with the present invention, effective production of measles virus which is a negative-strand RNA virus of the family *Paramyxoviridae*, is only obtained if the replicons specified by said cDNA molecule consist of an integral multiple of six nucleotides. This phenomenon will also be referred to as the "rule of six" throughout this application. The cDNA molecules can conveniently be used for the rescue of negative strand RNA viruses of the family *Paramyxoviridae*.

[0015] In a preferred embodiment of the present invention, said expression control sequence is an RNA polymerase promoter.

[0016] In another preferred embodiment of the present invention, said cDNA molecule is comprised in a plasmid. The plasmid is capable of propagation and preferably also expressing said cDNA molecule as an antigenomic RNA.

[0017] In a more preferred embodiment, said plasmid contains an expressible DNA fragment which replaces a preferably homologous DNA region of said cDNA molecule, or provides additional genetic information.

[0018] As was also found in accordance with the present invention, in the case of MV-based replicons the rule of six must be obeyed, if a foreign - homologous or heterologous - expressible DNA fragment is inserted into the plasmid containing said cDNA. In other words, any newly created replicon specified by appropriately constructed cDNA molecules will only be capable of yielding reasonable amounts of the desired product, if it obeys the rule of six.

[0019] In a most preferred embodiment, said plasmid is characterised in that the expressible DNA fragment is inserted into or adjacent to a region of said cDNA encoding a viral protein, said insertion being effected in a manner maintaining the reading frame to create a fusion protein and permitting the expression of said DNA fragment under the control of the signal sequences of said viral protein. In accordance with the present invention it is anticipated that in various cases appropriate C-terminal extensions of viral proteins will not interfere with their functionality.

[0020] In variation to the above described preferred embodiment and also comprised by the present invention, the expressible DNA fragment is expressed in such a manner downstream of a viral protein coding region to avoid formation of a fusion protein, but nevertheless allowing expression of the downstream coding sequence either by a stop/restart mechanism where the last A residue of the upstream termination triplet coincides with that of the start codon of the downstream coding region, or by placing an internal ribosome entry site (IRES) between the two coding regions; see example 12, second paragraph.

[0021] In a further most preferred embodiment, said plasmid is characterised in that the expressible DNA fragment is inserted into a non-coding region of said cDNA and flanked by viral signal sequences or heterologous signal sequences controlling the expression of the RNA fragment specified by said DNA fragment; see example 12, first paragraph.

[0022] Examples of this embodiment, creating additional transcription units, are provided by the plasmids specifying MVs expressing the heterologous CAT reading frame shown in Figure 10.

[0023] In a further more preferred embodiment of the invention, said plasmid comprises a genomic ribozyme sequence immediately adjacent to the 3' terminal nucle-

otide of said cDNA molecule and optionally downstream of said genomic ribozyme sequence at least one terminator, preferably the T7 terminator.

[0024] The inclusion of a ribozyme sequence into the plasmid of the invention leads to the faithful cleavage of the RNA transcript, thus greatly enhancing the yield of transcripts bearing the correct 3' termini which, in the case of MV, must obey the rule of six.

[0025] In a most preferred embodiment, said genomic ribozyme sequence is the hepatitis delta virus genomic ribozyme sequence.

[0026] In a further more preferred embodiment, said plasmid bearing said cDNA is capable of replicating in a prokaryotic host. A preferred example of such a prokaryotic host is *E. coli*. Illustrations of this preferred example are all cDNA constructs giving rise to modified MVs as shown in Figures 2 and 10 demonstrating plasmids replicating to high copy number in *E. coli*.

[0027] Additionally, in a more preferred embodiment said plasmid bearing said cDNA(s) is capable of replicating in a eukaryotic host.

[0028] The invention envisages the replication and expression (i.e. transcription, followed by translation of the transcripts formed) of the rescued vector, i.e. the packaged RNA particles (RNPs), in any suitable eukaryotic, preferably vertebrate, host cell. Preferred host cells are those with a high replication and expression capacity. Most preferred are those host cells that allow an easy recovery of rescued viruses for further replication and subsequent formulation in vaccines.

[0029] In another more preferred embodiment said expressible DNA fragment of said plasmid is a DNA fragment being homologous or heterologous with respect to the negative-strand RNA virus and encoding at least one immunogenic epitope.

[0030] In a most preferred embodiment of the present invention, in said plasmid said expressible DNA fragment encodes at least one immunogenic epitope of at least one pathogen, preferably an envelope protein, at least one gene product lacking in genetically defective individuals or toxic for targeted malignant cells.

[0031] This most preferred embodiment of the invention allows for the construction of plasmids as a basis for vaccines that effectively induce an immune response against one or preferably various different pathogens. In the case that the expressible DNA fragment encodes an envelope protein of a different virus than measles virus or of another pathogen, a measles virus based plasmid can be used to target specific cell types usually not recognised by measles virus. Said cell types can then selectively be targeted by rescued viruses specified by the plasmid of the invention and confer to said cell type, for example, a molecule that said cell type is in need of or a toxin, if said cell type is to be eliminated. Naturally, said molecule or toxin is also to be encoded by said plasmid. The person skilled in the art is capable of devising further applications of this basic principle for which the plasmid of the invention can be used.

[0032] Also, said plasmid can encode a product lacking in genetically defective individuals. The rescued virus can then be used for gene therapy of said genetically defective individuals.

[0033] Further, malignant cells can be targeted by the rescued virus which is based on said plasmid and molecules toxic for said malignant cells may be delivered.

[0034] In a further most preferred embodiment of the present invention, in said plasmid said expressible DNA fragment is derived from a virus, a bacterium, or a parasite.

[0035] In a further preferred embodiment of the invention, said expressible DNA fragment of said plasmid encodes an immunogenic epitope being capable of eliciting a protective immune response.

[0036] In a further preferred embodiment, said cDNA molecule or plasmids are based on an RNA virus which is measles virus or mumps virus.

[0037] The invention further contemplates for its practice a prokaryotic or eukaryotic host cell transformed with a plasmid as described above. Preferred host cells have been discussed above.

[0038] The invention can be put into effect using a helper cell capable of expressing an RNA replicon from a cDNA molecule as described above; said cDNA molecule being comprised in the plasmid described above or a plasmid comprising a cDNA molecule for the production of negative-strand RNA virus of a family of the order *Mononegavirales* which is not a member of the family of the *Paramyxoviridae*, said cDNA molecule comprising the entire (+)-strand sequence, operatively linked to an expression control sequence, and optionally an expressible DNA fragment which replaces a preferably homologous DNA region of said cDNA molecule or provides additional genetic information, said expressible DNA fragment encoding preferably at least one immunogenic epitope of at least one pathogen, which most preferably is capable of eliciting a protective immune response, said cell further being capable of expressing proteins necessary for transcription, encapsidation and replication of said RNA.

[0039] Apart from the features described above, the cDNA molecule for the production of negative-strand RNA virus of a family of the order *Mononegavirales* which is not a member of the family of the *Paramyxoviridae* may also have in certain embodiments the characteristics of the cDNA molecules that were discussed herein above, optionally in conjunction with the plasmids described above.

[0040] In view of the problems the prior art was confronted with for rescuing non-segmented negative-strand RNA viruses, in accordance with the present invention paradigmatic cell lines providing as helper functions T7 RNA polymerase and MV N and P protein were developed. Rescue of MVs can be directly monitored after transfection with plasmids specifying antigenomic RNAs and MV L mRNA. In principle, analogous helper cell lines can be generated for any of these viruses; thus

this rescue approach is applicable for all *Mononegavirales* replicating in vertebrate cells.

[0041] Thus, in a preferred embodiment of the invention, said proteins necessary for encapsidation, transcription and replication of said RNA are an RNA polymerase, preferably T7 RNA polymerase and optionally T3 RNA polymerase, and N and P protein, preferably of the virus to be rescued. In accordance with the present invention, said proteins are expressed from stably transfected expression plasmids, henceforth defined as genomic expression.

[0042] Since the rescue system now developed, in contrast to the one used for rescue of RV (Schnell et al., 1994), VSV (Lawson et al., 1995) and very recently also for SeV (D. Kolakofsky, personal communication), does not rely on any helper virus, there is no need to separate the rescued virus from the vast excess of any helper virus. Elimination of vaccinia virus from rescued virus is accomplished by a simple filtration step in the case of the rigidly structured virions of *Rhabdoviridae* but would involve more complex purification schemes in case of pleomorphic *Paramyxoviridae*, particularly those not replicating to high titers such as MV. Furthermore, for viruses impaired in replication and/or budding by the vaccinia virus, rescue using the prior art systems might fail altogether. Another possible drawback of the prior art systems based on the vaccinia helper virus is the high frequency of DNA recombinations occurring in the cytoplasm of vaccinia virus infected cells which might cause recombination of the plasmid bearing the antigenomic sequence with the plasmids encoding N, P and L protein required for the helper function; this may lead to rescue of viruses containing N, P and L sequences derived in part from the helper plasmids rather than from the plasmid bearing the antigenomic sequence. The helper cell system circumvents all of these problems and should in principle be applicable for the rescue of any of the *Mononegavirales* replicating in vertebrate cells.

[0043] It may not be necessary for the rescue of any single representative of *Mononegavirales*, to establish a helper cell line expressing the cognate N and P protein (in addition to T7 polymerase). Mini-replicon constructs containing the noncoding terminal regions (NCTs) of canine distemper virus (CDV) which is like MV a morbillivirus, differing from MV in 35% of the nucleotides in the NCTs, replicate in the MV-specific helper cells at an efficiency approaching that of the homologous MV mini-replicon. Thus, possibly CDV could be rescued with the 293-3-46 cells, which were developed in accordance with the present invention and more generally, any helper cell line might be able to rescue a number of not too distantly related *Mononegavirales*. This will probably depend on the compatibility of the proteins elicited by the related viruses, which was shown not to be the case for SeV-specific N and P and PIV3-specific L (Curran and Kolakofsky, 1991).

[0044] For the establishment of new helper cell lines

for other viruses which are also envisaged by the present invention, the following considerations might be helpful. The constitutive expression of the T7 RNA polymerase and the MV proteins N and P did not impair the long term stability of the 293-3-46 cell line, as mentioned in the examples attached hereto. Thus, inducible expression of these proteins, for example, by the approaches described by the group of Bujard (for a review see Gossen et al., 1993) will probably not be necessary, although it cannot be excluded that the N and P proteins of other viruses are more deleterious for cell growth than those of MV. Titration of the plasmids used for transfection proved essential, showing that a ratio of about 1:1000 of L-encoding and antigenome-producing plasmid, respectively, was optimal, in agreement with the deleterious effect of high VSV L expression for VSV replication noted by Schubert et al. (1985). An alternative mode of transiently supplying L, using a plasmid containing a CMV promoter/enhancer and an intron upstream rather than downstream of the L coding region to permit some export of the long L mRNA from the nucleus, was also successful in rescue, but the efficiency was not better than with the standard method of cytoplasmic T7-dependent L expression and more than a hundred times more L encoding plasmid was optimal for rescue. In view of these experiences, the decision not to include an L encoding plasmid for the generation of helper cells, thus allowing expression of L at adjustable ratios, was probably advantageous. Nevertheless, it should be mentioned that a cell line stably expressing SeV-derived N, P and L which mediates long term replication of natural SeV DIs has been described (Willenbrink and Neubert, 1994). It is important to note that this cell line differs fundamentally from the helper cells defined in the present invention by its lack of T7 polymerase. As a consequence, no rescue of a virus and not even of a minireplicon from cloned DNA is feasible with this cell line.

[0045] In a further preferred embodiment, said helper cell is transfected with at least one of said above described plasmids, said plasmids containing variant antigenomic cDNA of a representative of the *Mononegavirales*, and is additionally stably transfected with a plasmid comprising DNA encoding the cognate viral L protein.

[0046] Thus, instead of selecting for a helper cell that also encodes per se the viral polymerase (L protein), said L protein is transfected into said helper cell on a different plasmid, preferably by cotransfection. Further, a skilled person using the teachings of the present invention is able to create a suitable helper cell line expression also L protein, in which case cotransfection is not necessary.

[0047] In a most preferred embodiment, the genes encoding said N, P and L proteins are derived from measles or mumps virus.

[0048] In a further most preferred embodiment, said helper cell is derived from the human embryonic kidney

cell line 293 (ATCC CRL 1573). A preferred example of such a cell is clone 293-3-46 described in the examples. [0049] The invention further relates to an infectious negative-strand RNA virus strain belonging to the order *Mononegavirales* obtained by the method of the invention.

[0050] It must be recalled that five years ago, in an erroneous account, MV rescue was reported by our laboratory (Ballart et al., 1990 and EP-A 0 440 219), using the same basic principle. At that time, the experiments were based on microinjection of initiation complexes, consisting of T7 RNA polymerase and plasmids specifying MV genomes or antigenomes, into a particular cell line containing defective but replicating MV genomes. However, the rescue by microinjection experiments, unfortunately carried out by only one collaborator, could not be repeated, and all purportedly rescued viruses did not contain the genetic tag, as summarised in a commentary to these extremely sad and devastating events (Aldhous, 1992). It is now clear that rescue of MV could not be expected with that experimental setup for several reasons, in particular due to additional nucleotides at both ends of the generated RNAs and due to a cloning mistake rendering the RNA incompatible with the rule of six (Calain and Roux, 1993; the present invention).

[0051] The rescue efficiency, in comparison to rescue of positive-strand RNA viruses (Perrotta and Been, 1990), is low, since only 1 to 6 out of 10^6 transfected cells, each exposed on average to about 2.5×10^5 molecules of antigenomic and 80 to 800 molecules of L-encoding plasmid, trigger the formation of syncytia. Nevertheless, in comparison with the rescue method described for RV and VSV, where about 2×10^7 cells are transfected to obtain one rescue event (Lawson et al., 1995; Schnell et al., 1994), the MV rescue compares well, particularly in view of the fact that the MV genome size is roughly 4.5 kb larger and thus in principle more difficult to rescue. Importantly, the low efficiency should not constitute a difficulty for the rescue of MV variants replicating only to titer levels even orders of magnitude lower than the Edmonston B strains, since the bottleneck of rescue is constituted most likely by an early event. It is important to note that on cells fixed at various times after transfection, immunofluorescence indicating H or M gene expression was monitored exclusively in syncytia and there was no indication that rescue was confined to single cells. When rescue is visible directly by syncytia formation, already thousand of progeny MV genomes have arisen; impaired and thus slowly replicating virus variants might not form visible syncytia initially, but should be revealed after splitting of the transfected cell culture or upon seeding onto fresh Vero cells.

[0052] In a preferred embodiment of the method of the invention, the ratio of the plasmid comprising the cDNA molecule comprising the entire (+)-strand sequence of said negative-strand RNA virus operatively linked to an expression control sequence, which allows the synthesis of anti-genomic RNA transcripts bearing the authen-

tic 3'-terminal, and the plasmid comprising DNA encoding the viral L protein is about 1000:1.

In accordance with the present invention it has been shown that the above ratio is optimal for transfection efficiency.

[0053] In further preferred embodiments of the method of the invention, said recovering is either directly effected from the transfected helper cell culture after syncytia formation or, after mixing of detached helper cells with any other cells competent of being infected and replicating the assembled RNA viruses.

[0054] The invention relates further to a vaccine comprising the RNA virus according to the invention which optionally is obtainable by the method of the invention described above, optionally in combination with a pharmaceutically acceptable carrier.

[0055] The advantages of the vaccine of the present invention will be briefly discussed below.

[0056] In the past, a variety of DNA viruses and positive-strand RNA viruses have been used as carriers to direct the expression of heterologous genes or gene segments in host cells, mainly with the aim to elicit immune protection against the pathogen from which the heterologous genetic material was derived. The main advantage of using such live vaccines is their ability to multiply and typically infect a variety of different cell types, generating the antigens of interest intracellularly which can therefore be presented efficiently to the immune system, thus facilitating the induction of both T cell help and cytotoxicity. In contrast, killed vaccines or proteins manufactured by recombinant DNA technology are much less efficient, even by administration in various particulate forms developed recently, which are more efficient than traditionally used adjuvants. In addition, such vaccines typically induce no mucosal immunity, which is very important for protection against pathogens entering by the respiratory or intestinal route. Failure to induce mucosal immunity is also typical for the immunisation approach using injection of naked DNA encoding antigens.

[0057] On the other hand, most replicating vaccines constitute a possible threat, even if they are not proliferating, such as avipox vectors in humans (Baxby and Padietti, 1992). Complex viral vectors (e.g. based on vaccinia virus and related pox viruses, adenoviruses of herpesviruses) and bacterial vectors (e.g. based on derivatives of the agents causing tuberculosis or cholera) inherently elicit many lateral, unnecessary and/or undesired immune responses. In addition, DNA integration in the genome of infected or transfected cells bears at least the potential for malignant transformation. Multiauthored assessments of various types of vaccines have been published recently (Vaccines and public health; Internat. J. of techn. Ass. in Health care 10, 1-196 1994; Science 265, 1371-1451, 1994), from which the particular benefits of small RNA-based live vaccines are evident.

[0058] Several engineered positive-strand RNA viruses

have been described for potential use as vectors for immunisation purposes; early examples include poliovirus (Burke et al., 1988) and Sindbis virus (Xiong et al., 1989) and among several more recent accounts, involving larger polypeptide fragments expressed from various representatives of the *Picornaviridae*, just one should be mentioned here (Andino et al., 1994).

[0059] However, it must be stressed that the use of RNA viruses as vectors for vaccination purposes crucially depends on the stability of the foreign genetic material during the replication of the virus. This is not a trivial problem, because these viruses rely on a polymerase devoid of proofreading activity. Said problem has advantageously been solved by the present invention: in comparison to vaccine vectors based on positive-strand RNA viruses as mentioned above, the vaccine of the invention as exemplified by MV-based di- or multivalent vaccines show several important advantages which are valid in principle for any other member of the *Paramyxoviridae* such as mumps virus. First, the size of inserts is not a priori limited by a requirement to fit into an icosahedral protein shell. Second, the tight encapsidation of the genomes of *Mononegavirales* obviates RNA secondary structure which is very important in case of the positive-strand RNA viruses over the whole genome length to allow proper replication without annealing of the product to the template RNA strand; RNA segments encoding foreign antigens are not evolved to meet such requirements. Third, due to the modular set up of the genome, different insertion sites and expression modes, either as additional transcription units or as elongation of existing transcription units, expressing the inserted downstream reading frames by stop/restart or by an internal ribosome entry site can be envisaged, thus allowing a large range of different expression levels according to the position within the MV transcription gradient. Fourth, due to extremely low recombination frequencies, *Mononegavirales* can be expected to retain non-essential genetic material much more stably than positive-strand RNA-viruses. Finally, the rule of six, valid for MV as was found in accordance with the present invention and for other *Paramyxovirinae* (Calain and Roux, 1993), but as judged from cognate mini- and midi-replicons, not for *Rhabdoviridae* (Conzelmann and Schnell, 1994) or for *Pneumovirinae* (Collins et al., 1993), should even increase the faithful retention of foreign coding regions inserted in *Paramyxovirinae* in comparison to other *Mononegavirales*. Such an additional genetic stability can be anticipated because only one in six adventitiously arising large deletions and no small insertion or deletion of 1 to 5 nucleotides in a region nonessential for viral replication are expected to lead to viable progeny.

[0060] Further, knowledge of the nucleotide sequence variants conferring attenuation will allow to change the coding sequences not implicated in attenuating properties according to the evolution of the viruses over the years thus permitting to "update" the vaccines without incurring the danger of losing the quality of at-

fenuation.

[0061] The invention additionally contemplates for its practise the use of the plasmid described above in somatic gene therapy.

[0062] Since viral envelope proteins can be exchanged among different representatives of *Mononegavirales*, as shown here by the replacement of the MV envelope proteins with the VSV glycoprotein, it seems feasible to target the replion based on the replication machinery of *Mononegavirales* to particular cell types; thus, certain applications in somatic gene therapy can be envisaged. Advantages in comparison to existing vectors or gene therapy include their small size, thus limiting antigen reactions to a few proteins, and their complete inability to integrate into DNA and thus to transform cells.

[0063] Additionally, the invention can be put into effect using the plasmid described above for targeting special cell types. An outline of such targeting schemes and applications has been provided above.

[0064] The invention may further find application in the use of the plasmid described above for the functional appraisal of mutations found typically in MV variants responsible for fatal subacute sclerosing panencephalitis or for the identification of mutations responsible for attenuation of Paramyxoviridae strains, preferably measles virus strains.

[0065] Finally, the invention can be practised through a diagnostic composition comprising at least one cDNA molecule and/or at least one plasmid as described above.

THE FIGURES SHOW:

[0066]

Figure 1: Genomic map of measles virus

Figure 2: Plasmid vectors specifying RNAs with correct MV-specific termini. The numbers below the plasmid names indicate the length in nucleotides of the RNAs generated after ribozyme self-cleavage. Genomic or antigenomic sense of the specified RNAs is indicated by (-) and (+), respectively. Note that the MV nucleotide sequences present in these plasmids deviate in 30 positions from EMBL accession No K01711, most notably by a deletion of an A residue at pos. 30, compensated by insertion of an A at pos. 3402. For a commented overview of a MV consensus sequence see Radecke and Billeter (1995).

Figure 3: Western blot showing the expression of MV N and P proteins in MV-infected 293 cells, uninfected 293 cells and in cell line clones 293-3-46 and 293-3-64, respectively. Ar-

rows indicate the position of the structural MV N and P proteins as well as the non-structural V protein arising from MV P gene transcript editing.

Figure 4: Overview of experimental components and procedures for rescue. A: Mini-replicon rescue, implicating transfection of in-vitro transcribed RNA and coinfection with MV, supplying helper proteins N, P and L (and for later stages also M, F and H, as well as non-structural proteins C and V). B: MV rescue, implicating transfection of plasmid DNAs into helper cells mediating both artificial T7 transcription and N and P functions. For explanation of most symbols see Figure 2. The L encoding plasmid pEMC-La contains an internal ribosome entry site derived from encephalomyocarditis virus (stippled oval, EMC IRES), fused to the L coding region such that the initiator AUG of EMCV and L coincide; a poly dA tract downstream (about 40 dAs) is indicated as pdA. These two devices ensure transcript stability as well as efficient translation from the transcripts generated in the cytoplasm.

Figure 5: Assay of CAT activity elicited in 293-3-46 helper cells by transfection of the plasmid constructs p107MV(-):CAT and p107MV(-):CAT, specifying mini-replicons, and construct p(+):NP:CAT, specifying a midi-replicon. The backbone of the plasmid pT7P2lacZ is similar as described in Peltier and Sonenberg (1988). The CAT reading frame of the original plasmid is replaced by the lacZ reading frame.

Figure 6: Visualisation of syncytia formed in 293-3-46 helper cells. A: Rescue experiment, viewed by phase contrast microscopy 4 days after transfection. B, C: Cells grown on glass cover slips, fixed 3 days after transfection and viewed by phase contrast (B) or indirect immunofluorescence microscopy using a monoclonal antibody directed against MV M protein (C). Similar results were obtained with an antibody against H. The bar length represents 100µm.

Figure 7: Sequence determination of plaque-purified viruses, carried out by RT-PCR followed by cycle sequencing as described in the Examples. The left lanes of the relevant area reproduced from a sequencing gel relate to our laboratory Edmonston B strain, the right lanes to the rescued virus. Nucleotide

positions indicated correspond to those in the MV consensus sequence as defined in Figure 2.

Figure 8: Replication behaviour of plaque-purified viruses, evaluated by an overlay technique as described in the Examples. The derivatives of rescue experiments, the standard MV tag EdB and the 504 nucleotide deletion mutant MVΔ5F EdB are compared with a clone from our laboratory Edmonston B virus strain. The results of two independent experiments using a representative clone of each virus species are shown.

Figure 9: Northern blots revealing mRNAs of the rescued MV derived from p(+)-MV, and the MV deletion mutant derived from p(+)-MVΔ5F (Figure 2). The monocistronic F, M and H mRNA species (open triangles) and the bicistronic MF and FH mRNAs (black triangles) are revealed by M, F, and H-specific probes. The F-specific mono- and bicistronic RNAs induced by the deletion mutant are clearly smaller than the corresponding RNAs induced by the rescued standard MV (ΔF, 1859 rather than 2372 nt. calculated, without considering poly A tails; ΔMF, 3338 rather than 3842 nt., and ΔFH, 3830 rather than 4334 nt.).

Figure 10: Plasmids for production of standard and deleted MVs and hybrid MVs containing additional genes or exchanged envelope proteins. Note that two MV chimeric clones recovered from p(+)-MPCATV and from p(+)-MHCATV after 10 cycles of infection still expressed CAT activity encoded by the additional transcription unit in every one of the 10 clones taken from the tenth cycle tested.

Figure 11: Electron microscopy of BHK cells infected with replicating agent rescued from p(+)-MGV. Large arrays of RNPs typical for MV-infected cells are visible, showing unimpaired replication capability of the chimeric viral RNA.

Figure 12: Electron microscopy of BHK cells infected with replicating agent rescued from p(+)-MGV. Pleomorphic particles resembling MV virions are formed despite the fact that in these infected cell cultures exclusively VSV G protein and no trace of the MV envelope proteins F and H was detectable by West-

ern blotting.

Figure 13: Electron microscopy of BHK cells infected with VSV. VSV virion particles. The typical bullet-shaped VSV virions differ completely from the pleomorphic MV-like particles shown in Fig. 12.

[0067] The examples illustrate the invention:

EXAMPLE 1: CELLS AND VIRUSES

Cells were maintained as monolayers in Dulbecco's modified Eagle's medium (DMEM) supplemented with 5% foetal calf serum (FCS) for Vero cells (African green monkey kidney), with 10% FCS for 293 cells (human embryonic kidney) and with 10% FCS and 1.2 mg/ml G418 for the stably transfected 293 derived cell clones.

To grow MV virus stocks reaching titers of about 10^7 pfu/ml, recombinant viruses were propagated in Vero cells, and the vaccine strain Edmonston B was grown in Vero or 293 cells. One round plaque-purification was carried out by transferring a syncytium to a 35 mm Vero cell culture which was expanded to a 175 cm² dish. Virus stocks were made from 175 cm² cultures when syncytia formation was pronounced. Cells were scraped into 3 ml of OptiMEM I (GIBCO BRL) followed by one round of freezing and thawing. The virus titrations were carried out on 35 mm Vero cell cultures. After 2-3 h of virus adsorption, the inoculum was removed and the cells were overlaid with 2 ml of DMEM containing 5% FCS and 1% SeaPlaque agarose. After 4-5 days, cultures were fixed with 1 ml of 10% TCA for 1 h, then UV-cross linked for 30 min. After removal of the agarose overlay, cell monolayers were stained with crystal violet dissolved in 4% ethanol, and the plaques were counted.

EXAMPLE 2: GENERATION OF CELL LINE 293-3-46

Before the transfection, all plasmids were linearized by digestion with SfiI and sterilised by ethanol precipitation. Cells were seeded into one 35 mm well for transfection during 13 h as described below. The transfection mix contained 5 µg of pSC6-N, 4 µg of pSC6-P, and 1 µg of pSC6-T7-NEO. Then, cells were washed once with 2 ml of phosphate buffered saline (PBS; 137 mM NaCl, 2.7 mM KCl, 8 mM Na₂HPO₄, 1.5 mM

KH₂PO₄), and DMEM containing 10% FCS was added. After 2 days in culture, the cells of the 35 mm well were splitted to two 75 cm² dishes, and selection under 1.2 mg/ml G418 was started changing the medium every second day. After ~2 weeks, the first clones of a total of ~100 clones were transferred to 5 mm wells. When a clone had expanded to a 21 mm - or 35 mm well, cells were seeded for screening. The expression of the MV N and P proteins was analysed by Western blotting (see also below) using ~1/3 to 1/10 of the total lysate of a confluent 21 mm well. To monitor the functionality of the T7 RNA polymerase, a 35 mm cell culture was transfected with 4 µg of pEMC-Luc (Deng *et al.*, 1991), and the luciferase activity in 1/125 of the cleared total lysate (Promega protocol; harvest 1 day after transfection) was measured in a luminometer. Clones expressing the MV N and P proteins comparable to the same number of 293 cells infected with MV and showing a T7 RNA polymerase activity as high as possible were chosen to test their performance in allowing MV DI RNAs to express CAT. Here, 5 µg of the plasmids p107MV(+):CAT, p107MV(-):CAT, or p(+):NP:CAT with or without 100 ng of pEMC-La were transfected. After 1 day, cells were lysed, and 1/4 of the cleared lysates was tested for CAT activity.

EXAMPLE 3: PLASMID CONSTRUCTIONS

All cloning procedures were basically as described in Sambrook *et al.* (1989). PCR amplifications were carried out using the proofreading *Pfu* DNA polymerase (Stratagene) and primers with a 3' terminal phosphorothioate bond instead of a phosphodiester bond (Skerra, 1992). DNA sequences of the synthetic oligonucleotides are given in lower case for non-MV nucleotides and in upper case for the MV nucleotides; sequences of relevant restriction endonuclease recognition sites are underlined. The construction of the plasmid p107MV(-):CAT can be found in Sidhu *et al.*, 1995. Plasmid p107MV(+):CAT is the analogue of the plasmid p107MV(-):CAT. The additional intercistronic region of p(+):NP:CAT that is similar to the N-P intergenic boundary was constructed by inserting (5'-ctgcgcctaccctccatcattgt-TATMAMACTTAGGAACCGGTC

CACACAGCGCGCCAGCCCAT-CAACgcgcatcgcgata-3', MV(+):1717-1782) and the internally complementary oligonucleotide into the *SpeI* site of the P gene. The PCR-amplified CAT coding region was inserted as depicted in Figure 2.

The description of the assembly of the first MV full length DNA, the source of MV nucleotides 2044-14937 in later versions of full length clones such as *pauT7MV(-)* (see below), is given in Balart *et al.*, 1990. The main features of the plasmid p(+):MV (Figure 2) are as follows: The T7 promoter allows the synthesis of the MV antigenomic RNA precisely starting with the first nucleotide. The genomic hepatitis delta virus ribozyme (δ) liberates upon self-cleavage the correct MV 3' terminal nucleotide. Directly downstream of the δ ribozyme, the T7 RNA polymerase terminator Tφ stops most of the transcribing polymerases. This ensures that adjacent sequences derived from the vector backbone will not interfere with the cleavage activity. The cloning of p(+):MV started by annealing two internally complementary oligonucleotides #191 (5'-ggggaaccatcgatgga-taagaatcgccgcgcaggtac-3') and #192 (5'-ctgcgcgcgcattcttaccatcgatggt-tccccgc-3') yielding a short polylinker that carries the restriction sites for *SadI*, *Clal*, *NotI*, and *KpnI*. This new polylinker replaced the *SadI*-*KpnI* fragment in pBloT7 derived from pBluescript KS(+) (Stratagene) containing the T7 promoter fused to a *NsiI* site (Kaelin, 1989) thus forming the plasmid pBloT7NSCNK. To clone in the 5'-terminal 2041 bp of the MV antigenome (up to the *SadI* site), a *NsiI*-digestion was followed by treatment with Klenow polymerase in the presence of all four dNTPs. This created a blunt-end cloning site flush to the nontranscribed part of the T7 promoter sequence. A MV fragment comprising the nucleotides 1-2078 was generated from the 3351 bp *PvuI*-fragment of *pauMV(-)* by PCR amplification using primers #182 (5'-ACCAAACAAAGTTGGGTAAGGATAG-3', MV(+):1-25), and #183 (5'-CAGCGTCGTCATCGCTCTCTCC-3', MV(-):2077-2056). Note that the additional A residue at position MV(+):30 (Sidhu *et al.*, 1995) derived from the MV sequence of *pauMV(-)* was later deleted by mutational PCR. Upon *SadI*-treat-

ment, the MV fragment was ligated into the vector to yield pT7MV(+). Next, the 3'-terminus of the antigenome was linked to the sequence of δ followed downstream by T ϕ . The MV 3'-fragment (nucleotides 14907-15894) was generated from the 14046 bp PvuI-fragment of *pauMV(-)* by PCR amplification using the primers #186 (5'-GAGAAGCTAGAG-GAATTGGCAGCC-3'; MV(+)) 14907-14930) and #187 (5'-tctgaagactcACCA-GACAAAGCTGGG-3', MV(-) 15894-15879). Another PCR amplification on the plasmid *pau3a5T ϕ* with the primers #184 (5'-ataagaatgcggccgcatccgatat-agttcctcc-3') and #FR4 (5'-tctgaagactcTGGTggccggcatgtccag-3', MV(+)) 15891-15894) yielded the genomic HDV ribozyme linked to the T ϕ . Both primers #FR4 and #187 contain close to their 5' ends the recognition sequence for BbsI which creates a sticky end on both fragments comprising the four 3'-terminal MV nucleotides (MV(+)) TGGT). After the digestions of the MV 3'-fragment with ClaI and BbsI, of the δ /T ϕ -fragment with BbsI and NotI, and of pT7MV(+)'s with ClaI and NotI, a three-way ligation yielded the plasmid pT7MV(+)'s 5'3' δ T ϕ . The final step to generate p(+)'s MV was to fill in the remaining antigenomic MV nucleotides 2044-14937 by a three-way ligation. The SacI-Pad fragment (MV(+)) nucleotides 2044-7242) and the Pad-ClaI fragment (MV nucleotides 7243-14937) were released from plasmid *pauT7MV(-)*. These two fragments were ligated into pT7MV(+)'s 5'3' δ T ϕ from which the remaining polylinker (SacI-ClaI) had been removed. The plasmid p(-)'s MV (Figure 2) was constructed similarly. The self-cleavage activity of δ was demonstrated by detecting the expected small 3' fragments of *in vitro* made RNAs on a 5% polyacrylamide/7M urea gel. To generate p(+)'s MV Δ 5F carrying a 504 nt-deletion (MV(+)) 4926-5429) in the 5' noncoding region of the F gene, first a PCR was carried out on plasmid pAeF1 (Huber, 1993) using primers #88 (5'-CcGAATCAAGACTCATCCAATGTCCATCATGG-3', MV(+)) 5430-5461) and #89 (5'-AGAGAGATTGCCCAATGGATTGACCG-3', MV(-)) 5550-5523). The PCR fragment digested with HpaI replaced the NarI-HpaI fragment in pAeF1. The NarI-Pad-fragment of this vector then

replaced the corresponding fragment in p(+)'s MV.

The vector backbone of pEMC-La is based on pTM1 (Moss *et al.*, 1990) in which a NcoI-site overlaps with an ATG trinucleotide. Using this ATG as the start codon, an open reading frame inserted into this NcoI-site is translationally controlled by the encephalomyocarditis (EMC) virus internal ribosome entry site (IRES). The MV L coding sequence linked to an artificial poly(dA)-tract was taken from vector pAeL (Huber, 1993) in two steps: first, a 405 bp fragment containing the MV nucleotides 9234-9630 was generated by PCR using primers #194 (5'-gtggatccATGGACTCGCTATC TGTC AACCC-3', MV(+)) 9234-9255) and #195 (5'-AGTTAGTGTCCCTTAAGCAT TGGAAAACC-3', MV(-)) 9630-9602); second, a 6265 bp fragment comprising nucleotides 9572-15835 of the MV L gene sequence joined to the poly(dA)-tract was excised with EcoRI. After removing the NcoI-EcoRI part of the polylinker in pTM1 and digesting the PCR fragment also with NcoI and EcoRI, a three-way ligation including the 6265 bp EcoRI-fragment yielded pEMC-La. To eliminate the T7 promoter located 5' of the CMV promoter/enhancer in the vectors pSC-N and pSC-P (Huber *et al.*, 1991), pSC6-N and pSC6-P were constructed by replacing a PvuI-EcoRI fragment with the corresponding fragment of pSP65 (Promega). pSC6-T7 was generated by exchanging the N gene insert of pSC6-N by the fragment carrying the T7 RNA polymerase gene of pAR 1173 (Davanloo *et al.*, 1984). pSC6-T7-NEO was constructed by ligation of the phosphoglycerol kinase promoter-neomycin-resistance cassette (Soriano *et al.*, 1991) into the unique AvrII site of pSC6-T7 using appropriate linker oligodeoxynucleotides. All cloning sites were verified by sequencing.

EXAMPLE 4: TRANSFECTION OF PLASMIDS AND HARVEST OF REPORTER GENE PRODUCTS

Cells were seeded into a 35 mm well to reach ~50-70% confluence when being transfected. 3-8 h before transfection, the medium was replaced with 3 ml of DMEM containing 10% FCS. G418 was omitted henceforth because of its toxic effect during transfection. All plasmids

were prepared according to the QIAGEN plasmid preparation kit. The protocol for the Ca^{2+} phosphate coprecipitation of the DNA was adapted from Rozenblatt *et al.* (1979). The plasmids (2-10 μg per 35 mm well) were diluted with 300 μl of 1x transfection buffer (137 mM NaCl, 4.96 mM KCl, 0.7 mM Na_2HPO_4 , 5.5 mM dextrose, 21 mM HEPES pH 7.03). 1 M CaCl_2 solution was added to a final Ca^{2+} -concentration of 125 mM, and the mix was incubated at 20°C for 30-120 min. The coprecipitates were added dropwise to the culture and the transfection was carried out at 37°C and 5% CO_2 for ~15 h. Then, the transfection medium was replaced with 3 ml of DMEM containing 10% FCS. The products of the reporter genes were harvested 24-37 h after transfection. Cells were washed and lysed with Reporter lysis buffer (Promega), and CAT and luciferase assays were done following the supplier's protocol.

EXAMPLE 5: EXPERIMENTAL SET-UP TO RESCUE MV

293-3-46 cells prepared for transfection as described above were transfected with 5 μg of the plasmid harbouring the MV antigenomic DNA in presence or absence of 1-100 ng of the plasmid specifying the MV L mRNA. First syncytia appeared about 2-3 days after transfection when the cells were still subconfluent. To allow syncytia formation to progress more easily, almost confluent cell monolayers of each 35 mm well were then transferred to a 75 cm^2 dish. When these cultures reached confluence, cells were scraped into the medium and subjected once to freezing and thawing. Cleared supernatants were used to infect monolayers of Vero cells either to grow virus stocks or to harvest total RNA for analysis.

EXAMPLE 6: RT-PCR, CYCLE SEQUENCING, NORTHERN BLOT, WESTERN BLOT, IMMUNOFLUORESCENCE

For RT-PCR followed by cycle sequencing, Vero cells were infected with cleared virus suspensions either harvested from rescue cultures or from later passages, and total RNA was isolated according to Chomczynski and Sacchi (1987). 2 μg of total RNAs were first hybridised with 10 pmol or 1 nmol of random hexamer primers by heating to 80°C for 1 min and then

quick-cooled on ice. Reverse transcriptions were carried out with 200 U of MMLV-RT (GIBCO BRL) in the presence of 1 mM dNTPs in a buffer containing 20 mM Tris-HCl pH 8.4, 50 mM KCl, 2.5 mM MgCl_2 , 0.1 mg/ml bovine serum albumin, and 1 U RNasin (Promega). The mixes were kept at 20°C for 10 min, incubated at 42°C for 1 h, and terminated by heating at 95°C for 10 min. 1/10 of the reaction volumes was used as templates for the PCR amplification with the primers #59 (5'-ACTCGGTATCACTGCCGAG-GATGCAAGGC-3', MV(+) 1256-1284) and #183 (5'-CAGCGTCGTCATCGCTCTCTCC-3', MV(-) 2077-2056). After 40 cycles, the 822 bp fragments were isolated using the QIAquick gel extraction kit (QIAGEN). The sequencing reactions were done according to the linear amplification protocol (Adams and Blakesley, 1991). Primer #76 (5'-ctaGCCTAC-CCTCCATCATTGTTATAAAAACTTA G-3', MV(+) 1717-1749) was used for the tag in the 5' noncoding region of the P gene and primer #6 (5'-ccggTTATAA CAATGATGGAGGG-3', MV(-) 1740-1722) for the tag in the 3' noncoding region of the N gene.

Total cellular RNA for Northern blot analysis was isolated from Vero cells using the TRI REAGENT® (Molecular Research Center, Inc.) and poly(A) RNA was purified using oligo(dT)₂₅-coated super paramagnetic polystyrene beads (Dynal) and a magnetic particle concentrator. The RNA was electrophoresed through a 1% agarose gel in 6% formaldehyde-containing running buffer and transferred to a Hybond-N⁺ membrane (Amersham) by capillary elution in 20x SSC. Filters were prehybridised at 42°C for 4 h. Hybridisation was performed overnight at 42°C in 50% (v/v) formamide, 1 M NaCl, 10% (w/v) dextran sulfate, 1% SDS, yeast tRNA (0.1 mg/ml) containing 2×10^6 c.p.m./ml of an [α -³²P] dATP-labeled DNA probe prepared with Prime-It II (Stratagene). The following DNA fragments were used for random priming: the 1.4 kb *Sal*-*Bam*HI fragment from pSC-M (Huber *et al.*, 1991), the 1.7 kb *Hpa*I-*Pst*I fragment from pCG-F, and the 1.6 kb *Sma*I-*Xba*I fragment from pSC-H (Huber *et al.*, 1991). pCG, a eukaryotic expression vector containing a SV40 origin of replication and a CMV promoter/enhancer, was constructed by

deletion of the L gene as well as the downstream β -globin splice site of pSC-L (Huber et al., 1991; Severne et al., 1988) and subsequent insertion of the β -globin splice site (from pSG5 Strata-gene) upstream of a new polylinker. The pCG-based plasmid pCG-F contains an insert consisting of the entire F gene. Filters were washed in 2x SSC at 20°C for 10 min and twice in 2x SSC, 1% SDS at 65°C for 30 min. Bands were visualised by autoradiography.

To analyse the expression of the MV N and P proteins by Western blotting, cells were washed with PBS and cytoplasmic extracts were prepared using 300 μ l lysis buffer (50 mM Tris-HCl pH 8, 62.5 mM EDTA, 1% NP-40, 0.4% deoxycholate, 100 μ g/ml phenylmethylsulfonyl fluoride, and 1 μ g/ml Aprotinin). About 1/60 of the total lysates was run on SDS-8%PAGE and blotted onto Immobilon-P membranes. As first antibodies, either the rabbit polyclonal anti-N antibody #179 (kindly provided by C. Oervell prepared according to standard procedures) in a 6000-fold dilution in TBST (10 mM Tris-HCl pH 7.2-8, 150 mM NaCl, 0.05% Tween 20) or the rabbit polyclonal anti-P antibody #178 (Oervell and Norrby, 1980) in a 3000-fold dilution in TBST was used. The second antibody was a swine anti-rabbit antibody coupled to horseradish peroxidase allowing the visualisation of the bands by the enhanced chemiluminescence kit (ECL™ Amersham Life Science, RPN 2106).

For immunofluorescence microscopy, 293-3-46 cells were seeded for a rescue experiment on 24 mm x 24 mm glass cover slips in 35 mm wells, cultured overnight and transfected, as described above. 3 days after transfection, cells were permeabilized with acetone:methanol (1:1) and indirect immunofluorescence was performed essentially as described (Hancock et al., 1990; Oervell and Norrby, 1980), except that PBS was supplemented with 1 mM MgCl₂ and 0.8 mM CaCl₂ and that p-phenylenediamine was omitted from the mountant. Viral M and H proteins were detected using mouse monoclonal anti-M-16BB2 and anti-H-129 antibodies (Sheshberadaran et al., 1983) and rabbit anti-mouse IgG [F(ab')₂] antibodies coupled to rhodamine (Pierce, 31666).

EXAMPLE 7: GENOMIC AND ANTIGENOMIC PLASMIDS SPECIFYING MINI-, MID-, AND FULL LENGTH REPLICONS

The plasmid constructs used in this study are shown in Figure 2. p107MV(-): CAT and p107MV(+): CAT specify genome- and antigenome-sense RNAs, respectively, in which all MV coding regions are precisely replaced by the CAT coding region. In MV-infected cells or in helper cells (see below), they give rise to mini-replicons and to capped and polyadenylated CAT mRNA comprising the 5'N and the 3'L noncoding region. p(+)NP:CAT, containing in addition also the MV N and P coding regions in their ordinary MV sequence context, gives rise to mid-replicons. Full length or partially deleted antigenomic or genomic RNAs are specified by p(+)MVΔ5F, p(+)MV and p(-)MV. For all these plasmids, transcription with T7 RNA polymerase yields RNAs bearing the authentic nucleotides of the viral genomic and antigenomic termini, respectively (Sidhu et al., 1995). Correct initiation was accomplished by direct fusion of the T7 promoter (devoid of its transcribed part) to the genomic and antigenomic sequence. Starting all transcripts with the MV-specific nucleotides ACC rather than the T7-specific GGG reduces the RNA yield by about one order of magnitude, as revealed by in vitro transcription studies using precursor plasmid constructs. To mediate formation of the correct MV 3' termini, the hepatitis delta virus genomic ribozyme sequence (Perotta and Been, 1990) was cloned immediately adjacent to the MV 3' terminal nucleotides; the introduction of T7 terminators increased the efficiency of self-cleavage.

EXAMPLE 8: HELPER CELLS STABLY EXPRESSING MV N AND P PROTEIN AS WELL AS T7 RNA POLYMERASE

The human embryonic kidney cell line 293 was chosen because it is highly permissive for MV. In addition, these cells can be efficiently transfected by the calcium phosphate coprecipitation method; 30 to 60% of the cells stained blue 24 hours after transfection with a plasmid encoding β -galactosidase.

Following cotransfection of 293 cells with pSC6-N, pSC6-P and pSC6-T7-NEO as described in the Examples, about 100 colonies were expanded un-

der neomycin selection. The expression of N and P was screened by Western blotting, and the activity of T7 RNA polymerase was evaluated by transfection with a reporter plasmid containing the firefly luciferase coding region under control of a T7 promoter. Many clones expressed high levels of P, but only few coexpressed N efficiently. Figure 3 shows N and P expression of two selected cell lines at levels comparable to that of MV-infected 293 cells; T7 RNA polymerase activity detected in clone 293-3-46 was among the highest of all clones whereas it was about 100 times lower in clone 293-3-64 which turned out not to rescue MV. A third cell line, 293-3-43, expressing the three proteins at levels comparable to 293-3-46 was also active in rescue.

The expression of the introduced genes did not reduce the susceptibility for MV infection. The helper cell line 293-3-46 principally used MV rescue, although growing at a rate 2-3 times slower in comparison to the parent 293 line, proved to be very stable and fully functional after more than 80 cell splittings at dilutions 1:4 to 1:8.

EXAMPLE 9: FROM MV MINI-REPLICON RESCUE USING HELPER MV TO MV RESCUE USING HELPER CELLS 293-3-46

The MV rescue system was developed stepwise, permitting to functionally test all components. On one side, MV-dependent rescue of mini- and later successively longer midi-replicons was ascertained by CAT reporter assays. Similarly, on the other side, the functionality of the 293-3-46 cells was compared to the MV-based help described before (Sidhu et al., 1995).

The mini-replicon rescue test is shown schematically in Figure 4A. Small transcripts from p107MV(-):CAT, p107MV(+):CAT (Sidhu et al., 1995) and later longer transcripts, e.g. generated from p(+):NP:CAT (Figure 2), behaved like mini- and midi-replicons, respectively. They were encapsidated, transcribed to produce CAT, replicated and packaged into virion particles to infect new cells. During the first 2 to 4 infection cycles, they massively amplified whereas in later cycles replication of both MV and the mini-replicons was curtailed, as observed for naturally occurring DI RNAs (Re, 1991).

Analyses of the amplified RNAs showed that the encapsidated replicons and the CAT transcripts contained the respective different MV-specific terminal regions (Sidhu et al., 1995). Most importantly, it turned out that for efficient function, the total number of nucleotides of the replicons had to be a multiple of six, a requirement - termed the rule of six - previously found essential for natural and slightly modified SeV DI RNAs of the copy-back type (Calain and Roux, 1993). Adherence to this rule was crucial for the construction of plasmids specifying a variety of mini- and midi-replicons such as those shown in Figure 2. This was also the case for full length clones. The helper function of stably transfected cell clones was tested with the set-up represented in Figure 4B, using however either plasmid p107MV(-):CAT, p107MV(+):CAT or p(+):NP:CAT (Figure 2) instead of p(+):MV. As shown in Figure 5, CAT activity arose in the transfected cells, although at levels considerably lower than in 293 cells infected with MV and cotransfected directly with mini- or midi-replicon RNA. The cotransfection of plasmid pEMC-La encoding the MV L protein was an absolute requirement. As expected, low background CAT activity was detected when the plus-sense mini-replicon construct was used. The two constructs containing only the CAT reading frame in the plus- and minus-sense elicited about equal amounts of CAT activity; the midi-replicon construct gave rise to roughly 100 times less CAT activity than the mini-replicon.

The transfection protocol was optimised in terms of maximal achievable CAT activity, using mini- and midi-replicon plasmids. Then, the full length constructs p(+):MV and p(-):MV were tested. About 10^6 cells contained in each 35 mm well were transfected and we estimate that about one tenth of these actually received full length as well as the L-encoding plasmids. Usually, following cotransfection of p(+):MV and pEMC-La, 1 to 6 syncytia developed after 2 to 3 days in each well. No syncytia were found when the latter was omitted or when the p(-):MV plasmid was used. The rescue experiments were carried out by different experimenters using different DNA preparations. The efficiency was slightly variable, but at least 30% of the transfected

wells revealed rescue. Figure 6 shows typical syncytia formed in these experiments, viewed either directly (phase contrast, 6A) or after fixation of cells grown on cover slips (phase contrast, 6B, or immunofluorescence of the same area, 6C).

EXAMPLE 10: CHARACTERISATION OF RESCUED MV

First, it had to be ascertained that the rescued MVs contained the genetic tag which had been introduced into the MV full length plasmid clones. The 3 nt tag indicated in Figure 2 originated from a variant 176 nt N/P noncoding gene boundary region (NCGB) recovered from the SSPE-derived MV replicating in IP-3-Ca cells (Ballart et al., 1990). Rescued viruses were amplified in Vero cells, either directly from the transfected cells or after plaque purification; the products recovered by reverse transcription followed by polymerase chain reaction (RT-PCR) were analysed by cycle sequencing. Figure 7 shows an example of these analyses, revealing the AG tag instead of CA in the Edmonston B strain passaged in our laboratory.

We did not analyse the entire sequence of rescued MVs to exclude any error introduced either during the assembly of the antigenomic plasmid clones or during T7 RNA polymerase transcription in the rescue step. However, major deleterious changes could be ruled out by analysing the replication behaviour of the rescued virus in comparison to that of the Edmonston B strain. Figure 8 shows that both the speed of replication as well as the final titers reached in repeated experiments were indistinguishable between single plaque-purified normal (MV EdB) and rescued (MV tag EdB) viruses. The apparent difference at day 1 after infection was not a consistent observation. Non-plaque-purified virus stocks gave similar results.

EXAMPLE 11: MV MISSING 504 NUCLEOTIDES IN THE F GENE 5' NONCODING REGION

As a first application of the reverse genetics system, we deleted 504 nucleotides, thus generating a shortened genome compatible with the rule of six mentioned above. This eliminated almost the entire F gene segment of the long enigmatic noncoding M/F NCGB

which is typical for MV and the other morbilliviruses, whereas the representatives of the other two genera of the subfamily *Paramyxovirinae*, paramyxovirus and rubulavirus, contain only a short NCGB. Remarkably, it was viable and moreover it replicated in cell culture at a rate indistinguishable from that of the Edmonston B and the rescued nondeleted MV strain (Figure 8, MVΔSF EdB). To determine the size of the F gene derived RNAs, the MV-specific mRNA induced by these plaque purified viruses was analysed, using probes specific for the F and for the M and H genes situated up- and downstream of F, respectively. Indeed, as shown in Figure 9, the F mRNA as well as the MF and FH bicistronic RNAs are consistently shorter in cells infected with the MVΔSF EdB variant.

Example 12: MVs expressing CAT activity

To explore the feasibility to express foreign proteins from engineered MV we inserted a CAT reading frame flanked by intercistronic regions into the MV antigenomic cDNA sequence; two positions were tested, on one hand between the N and the P and on the other hand between the H and the L gene (Figure 10, p(+)MPCATV and p(+)MHCATV, respectively). The intercistronic region flanking the CAT reading frame was devised according to the intercistronic N/P gene boundary region, but contains additional restriction sites unique in the entire plasmid, suitable for further manipulations. From these constructs, recombinant MVs expressing CAT activity were rescued with about the same efficiency as from the standard and the deleted constructs p(+)MV and p(+)MΔSFV, respectively. As expected from the natural transcription gradient typical for all *Mononegavirales*, p(+)MHCATV expressed somewhat less CAT activity than p(+)MPCATV. Most importantly, the CAT expression of the recombinant viruses seems to be remarkably stable as revealed from the experiment mentioned in the legend to Figure 12 in which an overall amplification of the recombinant viruses of at least 10^{30} was achieved. We actually had expected that viruses rescued from p(+)MPCATV would be less stable than those from p(+)MHCATV, because in the former the transcription of all genes following the insert

ed CAT are expected to be lower than normal whereas in the latter only the L gene transcription should be lower. Apparently, the position of the insert does not greatly affect the viability of the rescued viruses. However, no competition experiments with standard MV have been carried out so far. Furthermore, it has to be expected that recombinant viruses expressing proteins which actively interfere with MV replication will turn out to maintain the inserted gene less faithfully.

It should be mentioned here that insertion of a foreign coding sequence within existing MV genes should be even less harmful for the viral replication than by creating new transcription units as in the constructs discussed above. The general inability of the eukaryotic translation machinery to express more than one reading frame from a mRNA can in principle be overcome by (at least) two devices: the stop/restart mechanism and internal ribosome entry sites (IRES). Both mechanisms are actually used in special cases for natural protein expression. An example of the first is represented by the translation of the M2 polypeptide in Influenza B virus (Horvath, C.M., Williams, M.A., and Lamb, R. A. (1990) Eukaryotic coupled translation of tandem cistrons; identification of the influenza B virus BM2 polypeptide. EMBO J. 9, 2639-2947). For the second mechanism, many recognized natural precedents exist, most notably the IRES of Picornaviridae (Sonenberg, N. (1990) Poliovirus translation. Curr. Top. Microbiol. Immunol. 161, 23-47), but also IRES in cellular mRNAs such as that specifying BiP (Samow, P. (1990) Translation of glucose-regulated protein 78/immunoglobulin heavy-chain binding protein mRNA is increased in poliovirus-infected cells at a time when cap-dependent translation of cellular RNA is inhibited). All of these cited types of device have been explored in the context of the MV N and H genes, using as coding regions downstream of the MV N and H reading frames those yielding CAT and firefly luciferase, respectively, as reporters. The whole bicistronic constructs were expressed from conventional expression plasmids in primate cells and yields of reporter proteins ranging between 10 and 100% in comparison to the proteins en-

coded by the upstream reading frames were obtained (Diploma theses, University of Zürich, composed by A. Cathomen (1991) and O. Peter (1992)).

Example 13: MV chimera bearing the VSV envelope protein

To explore the feasibility to rescue genetically stable chimeric *Mononegavirales* in which the envelope proteins of one virus are replaced by the those of another virus p(+)MGV and pMG/FV (Figure 10) were constructed. In the former construct the entire MV F and H coding regions were replaced by that encoding the VSV G protein which fulfills a receptor binding and a fusion function analogous to those of the MV H and F proteins, respectively. The latter construct was devised such that a fusion protein is created containing the large exterior part and the transmembrane region from the VSV G protein fused to the cytoplasmic tail of the MV F protein which is thought to interact specifically with the MV M protein. Indeed, chimeric viruses could be recovered from both constructs which could be distinguished from each other only by slightly different cytopathic effects (which are both drastically different from those elicited by MV) and by the fact that in cells infected by the virus rescued from the latter construct the fusion protein could be revealed by Western blotting not only by antibodies directed to the VSV G exodomain by also to antibodies directed against the MV F cytoplasmic tail. Both chimera replicated, as determined by end point dilutions, to reasonably high titers only about one order of magnitude lower than the titers obtained by MV. In addition, they showed the biological specificities expected: they readily infect rodent cells (which do not express a MV receptor) such as BHK (Figures 11, 12) where they form abundant cytoplasmic and nuclear RNPs typical for MV (Figure 11) as well as pleomorphic particles resembling MV virions (Figure 12) completely different from the tight shell- or cigar-like VSV virions (Figure 13) thought to be shaped primarily by the VSV M protein.

Considering the fact that MV and VSV are only very distantly related *Mononegavirales* and indeed belong to different families (*Paramyxoviridae* and *Rhabdoviridae*, respectively), it seems quite

likely that many different chimera involving more closely related *Mononagavirales* can be created and it appears not unrealistic that also chimera containing envelope proteins targeting particular cell receptors can be developed.

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Claims

1. A method for the production of an infectious non-segmented negative-strand RNA virus of the order *Mononegavirales* comprising
 - (a) introducing a cDNA molecule comprising the entire (+)-strand sequence of said negative-strand RNA virus operatively linked to an expression control sequence, which allows the synthesis of antigenomic RNA transcripts bearing the authentic 3'-termini, into a helper cell expressing an RNA-polymerase, preferably T7 RNA-polymerase, an N and a P protein, preferably of the virus to be rescued, and, further, an L protein, preferably of the virus to be rescued, encoded by a cDNA comprised by a plasmid either transiently or stably introduced into said cell; and
 - (b) recovering the assembled infectious non-segmented negative-strand RNA virus.
2. The method of claim 1 wherein said virus belongs to the family *Paramyxoviridae*.
3. The method of claim 1 or 2, wherein the expression control sequence of 1(a) is an RNA polymerase promoter.
4. The method of any one of claims 1 to 3, wherein said cDNA molecule is comprised in a plasmid.
5. The method of claim 4, wherein said plasmid contains an expressible DNA fragment which replaces a homologous DNA region of said cDNA molecule.
6. The method of claim 4, wherein said plasmid contains an expressible DNA fragment which provides additional genetic information.
7. The method of claim 4, wherein said plasmid contains an expressible DNA fragment which replaces a heterologous DNA region of said cDNA molecule.
8. The method of any one of claims 5 to 7, wherein said plasmid is characterized in that the expressible DNA fragment is inserted into a region of said cDNA encoding a viral protein, said insertion being effected in a manner maintaining the reading frame, preferably to create a fusion protein, and permitting the expression of said DNA fragment under the control of the signal sequences of said viral protein.
9. The method of claim 5 to 7, wherein said plasmid is characterized in that the expressible DNA fragment is expressed in such a manner downstream of a viral protein coding region to avoid formation of a fusion protein, but nevertheless allowing expression of the downstream coding sequence either by a stop/restart mechanism where the last A residue of the upstream termination triplet coincides with that of the start codon of the downstream coding region, or by placing an internal ribosome entry site (IRES) between the two coding regions.
10. The method of any one of claims 5 to 7, wherein said plasmid is characterized in that the expressible DNA fragment is inserted into a non-coding region of said cDNA and flanked by viral signal sequences or heterologous signal sequences controlling the expression of the RNA fragment specified by said DNA fragment.
11. The method of any one of claims 4 to 10, wherein said plasmid comprises a genomic ribozyme sequence immediately adjacent to the 3' terminal nucleotide of said cDNA molecule and optionally downstream of said genomic ribozyme sequence at least one terminator, preferably the T7 terminator.
12. The method of claim 11, wherein said genomic ribozyme sequence is the hepatitis delta virus genomic ribozyme sequence.
13. The method of any one of claims 4 to 12, wherein said plasmid is capable of replicating in a prokaryotic or eukaryotic host.
14. The method of any one of claims 5 to 13, wherein said expressible DNA fragment of said plasmid is a DNA fragment being homologous or heterologous

with respect to the negative-strand RNA virus and encoding at least one immunogenic epitope.

15. The method of claim 14, wherein said expressible DNA fragment of said plasmid encodes at least one immunogenic epitope of at least one pathogen, preferably an envelope protein, or at least one gene product lacking in genetically defective individuals or toxic for targeted malignant cells. 5
16. The method of claim 15, wherein said expressible DNA fragment of said plasmid is derived from a virus, a bacterium, or a parasite. 10
17. The method of any one of claims 5 to 16, wherein said expressible DNA fragment of said plasmid encodes an immunogenic epitope being capable of eliciting a protective immune response. 15
18. The method of any one of claims 2 to 17, wherein said negative-strand RNA virus is measles virus or mumps virus. 20
19. The method of any one of claims 1 to 17, wherein in said helper cell said N, P and L protein encoding genes are derived from measles or mumps virus. 25
20. The method of any one of claims 1 to 19, wherein said helper cell is derived from the human embryonic kidney cell line 293 (ATCC CRL 1573). 30
21. The method of any one of claims 1 to 20, wherein the ratio of the plasmid as defined in any one of claims 4 to 20 and the plasmid comprising DNA encoding the viral L protein is about 1000:1. 35
22. The method of any one of claims 1 to 21, wherein said recovery of said virus is achieved directly from the transfected helper cell culture after syncytia formation. 40
23. The method of any one of claims 1 to 22, wherein said recovery of said virus is achieved after mixing of the transfected helper cell with other cells competent of being infected and capable of replicating said virus. 45
24. An infectious non-segmented negative-strand RNA virus of the family *Paramyxoviridae* obtained by the method of any one of claims 7 to 10. 50
25. A vaccine comprising the RNA virus of claim 24, optionally in combination with a pharmaceutically acceptable carrier. 55

Patentansprüche

1. Verfahren zur Herstellung eines infektiösen nicht-segmentierten Negativ-Strang-RNA-Virus der Ordnung *Mononegavirales*, umfassend:
 - (a) Einführung eines cDNA-Moleküls, umfassend die gesamte (+)-Strangsequenz des Negativ-Strang-RNA-Virus, funktionell mit einer Expressionskontrollsequenz verbunden, welche die Synthese von die authentischen 3'-Termini umfassenden antigenomischen RNA-Transkripten ermöglicht, in eine Helferzelle, die eine RNA-Polymerase exprimiert, vorzugsweise T7 RNA-Polymerase, ein N- und ein P-Protein, vorzugsweise des zu gewinnenden Virus, und des weiteren ein L-Protein, vorzugsweise des zu gewinnenden Virus, kodiert von einer in einem Plasmid enthaltenen cDNA, das entweder transient oder stabil in die Zellen eingeführt worden ist; und
 - (b) Gewinnung des assemblierten infektiösen nicht-segmentierten Negativ-Strang-RNA-Virus.
2. Verfahren nach Anspruch 1, wobei der Virus zu der Familie der *Paramyxoviridae* gehört.
3. Verfahren nach Anspruch 1 oder 2, wobei die Expressionskontrollsequenz von 1(a) ein RNA-Polymerase-Promotor ist.
4. Verfahren nach einem der Ansprüche 1 bis 3, wobei das cDNA-Molekül in einem Plasmid enthalten ist.
5. Verfahren nach Anspruch 4, wobei das Plasmid ein exprimierbares DNA-Fragment enthält, welches eine homologe DNA-Region des cDNA-Moleküls ersetzt.
6. Verfahren nach Anspruch 4, wobei das Plasmid ein exprimierbares DNA-Fragment enthält, das zusätzliche genetische Informationen bereitstellt.
7. Verfahren nach Anspruch 4, wobei das Plasmid ein exprimierbares DNA-Fragment enthält, das eine heterologe DNA-Region des cDNA-Moleküls ersetzt.
8. Verfahren nach einem der Ansprüche 5 bis 7, wobei das Plasmid dadurch gekennzeichnet ist, daß das exprimierbare DNA-Fragment in eine Region der cDNA eingefügt ist, die ein virales Protein kodiert, wobei die Insertion in einer Weise ausgeführt ist, die das Leseraster beibehält, vorzugsweise um ein Fusionsprotein herzustellen, und die Expression des DNA-Fragments unter der Kontrolle der Signalsequenzen des viralen Proteins ermöglicht.

9. Verfahren nach einem der Ansprüche 5 bis 7, wobei das Plasmid dadurch gekennzeichnet ist, daß das exprimierbare DNA-Fragment derart stromabwärts von einer viralen protein-kodierenden Region exprimiert wird, daß die Bildung eines Fusionsproteins vermieden, aber nichtdestotrotz die Expression der stromabwärts befindlichen kodierenden Sequenz ermöglicht wird, entweder durch einen Stop-/Neustart-Mechanismus, wobei der letzte A-Rast des stromaufwärts liegenden Terminationstriplets mit demjenigen des Startkodons der stromabwärts liegenden kodierenden Region übereinstimmt, oder durch das Platzieren einer internen Ribosomeintrittsstelle (IRES) zwischen die beiden kodierenden Regionen.
10. Verfahren nach einem der Ansprüche 5 bis 7, wobei das Plasmid dadurch gekennzeichnet ist, daß das exprimierbare DNA-Fragment in eine nicht kodierende Region der cDNA eingefügt ist und durch virale Signalsequenzen oder heterologe Signalsequenzen flankiert wird, die die Expression des durch das DNA-Fragment bestimmten RNA-Fragments kontrollieren.
11. Verfahren nach einem der Ansprüche 4 bis 10, wobei das Plasmid eine genomische Ribozymsequenz aufweist, die unmittelbar benachbart ist zum 3'-terminalen Nukleotid des cDNA-Moleküls, und gegebenenfalls stromabwärts der genomischen Ribozymsequenz mindestens einen Terminator, vorzugsweise den T7-Terminator.
12. Verfahren nach Anspruch 11, wobei die genomische Ribozymsequenz die Hepatitis-delta-Virus genomische Ribozymsequenz ist.
13. Verfahren nach einem der Ansprüche 4 bis 12, wobei das Plasmid in einem prokaryotischen oder eukaryotischen Wirt replizieren kann.
14. Verfahren nach einem der Ansprüche 5 bis 13, wobei das exprimierbare DNA-Fragment des Plasmids in Bezug auf das Negativ-Strang-RNA-Virus ein homologes oder heterologes DNA-Fragment ist und mindestens ein immunogenes Epitop kodiert.
15. Verfahren nach Anspruch 14, wobei das exprimierbare DNA-Fragment des Plasmids mindestens ein immunogenes Epitop von mindestens einem Pathogen kodiert, vorzugsweise von einem Hüllprotein, oder mindestens ein Genprodukt, das in genetisch defekten Individuen fehlt oder toxisch für angreifene maligne Zellen ist.
16. Verfahren nach Anspruch 15, wobei das exprimierbare DNA-Fragment des Plasmids von einem Virus, einem Bakterium oder einem Parasiten stammt.
17. Verfahren nach einem der Ansprüche 5 bis 16, wobei das exprimierbare DNA-Fragment des Plasmids ein immunogenes Epitop kodiert, das eine schützende Immunantwort hervorrufen kann.
18. Verfahren nach einem der Ansprüche 2 bis 17, wobei der Negativ-Strang-RNA-Virus Masern-Virus oder Mumps-Virus ist.
19. Verfahren nach einem der Ansprüche 1 bis 17, wobei die in der Helferzelle das N-, P- und L- Protein kodierenden Gene vom Masern- oder Mumps-Virus stammen.
20. Verfahren nach einem der Ansprüche 1 bis 19, wobei die Helferzelle von der menschlichen embryonischen Nierenzelllinie 293 (ATCC CRL 1573) stammt.
21. Verfahren nach einem der Ansprüche 1 bis 20, wobei das Verhältnis des in einem der Ansprüche 4 bis 20 definierten Plasmids und des Plasmids, welches DNA beinhaltet, die das virale L-Protein kodiert, ungefähr 1000:1 ist.
22. Verfahren nach einem der Ansprüche 1 bis 21, wobei die Gewinnung des Virus direkt von der transfizierten Helferzellkultur nach Syncytien-Formation erfolgt.
23. Verfahren nach einem der Ansprüche 1 bis 22, wobei die Gewinnung des Virus nach Vermischen der transfizierten Helferzelle mit anderen Zellen erfolgt, die infektiionskompetent sind und das Virus replizieren können.
24. Infektioser nicht-segmentierter Negativ-Strang-RNA-Virus der Familie *Paramyxoviridae*, der durch das Verfahren nach einem der Ansprüche 7 bis 10 erhalten wird.
25. Impfstoff umfassend den RNA-Virus von Anspruch 24, gegebenenfalls in Kombination mit einem pharmazeutische verträglichen Träger.

Revendications

1. Procédé pour la production d'un virus ARN à brin négatif non segmenté infectieux de l'ordre Mononegavirales, comprenant les étapes consistant à :
 - (a) introduire une molécule ADNc comprenant toute la séquence (+)-brin dudit virus ARN brin négatif lié de façon opérante à une séquence de commande d'expression qui permet la synthèse des produits de transcription ARN antigénomiques portant les terminaisons 3'

- authentiques, dans une cellule auxiliaire exprimant une polymérase ARN, de préférence une polymérase ARN T7, une protéine N et une protéine P, de préférence du virus à récupérer et de plus, une protéine L, de préférence du virus à récupérer, codé par un ADNc compris dans un plasmide introduit de façon transitoire ou stable dans ladite cellule; et
(b) récupérer le virus ARN infectieux à brin négatif non segmenté assemblé.
2. Procédé selon la revendication 1, dans lequel ledit virus fait partie de la famille des Paramyxoviridae.
 3. Procédé selon la revendication 1 ou 2, dans lequel la séquence de commande d'expression de 1(a) est un promoteur de polymérase ARN.
 4. Procédé selon l'une des revendications 1 à 3, dans lequel ladite molécule ADNc est comprise dans un plasmide.
 5. Procédé selon la revendication 4, dans lequel ledit plasmide contient un fragment ADN exprimable qui remplace une région ADN homologue de ladite molécule ADNc.
 6. Procédé selon la revendication 4, dans lequel ledit plasmide contient un fragment ADN exprimable qui fournit des informations génétiques supplémentaires.
 7. Procédé selon la revendication 4, dans lequel ledit plasmide contient un fragment ADN exprimable qui remplace une région ADN hétérologue de ladite molécule ADNc.
 8. Procédé selon l'une quelconque des revendications 5 à 7, dans lequel ledit plasmide est caractérisé en ce que le fragment d'ADN exprimable est inséré dans une région dudit ADNc codant une protéine virale, ladite insertion étant effectuée d'une manière qui maintient le cadre de lecture, de préférence pour créer une protéine de fusion et permettant l'expression dudit fragment d'ADN sous le contrôle des séquences de signaux de ladite protéine virale.
 9. Procédé selon la revendication 5 à 7, dans lequel ledit plasmide est caractérisé en ce que le fragment d'ADN exprimable est exprimé en aval d'une région codant la protéine virale de manière telle à éviter la formation d'une protéine de fusion, mais permettant néanmoins l'expression de la séquence codant en aval soit par un mécanisme arrêt/repêchage lorsque le dernier résidu A du triplet de terminaison amont coïncide avec celui du codon de départ de la région codant en aval soit en plaçant un site d'entrée de ribosome interne (SRI) entre les deux régions codantes.
 10. Procédé selon l'une quelconque des revendications 5 à 7, dans lequel le plasmide est caractérisé en ce que le fragment d'ADN exprimable est inséré dans une région non codante dudit ADNc et flanqué des séquences de signaux viraux ou des séquences de signaux hétérologues commandant l'expression du fragment ARN spécifié par ledit fragment d'ADN.
 11. Procédé selon l'une quelconque des revendications 4 à 10, dans lequel ledit plasmide comprend une séquence ribozyme génomique immédiatement contiguë au nucléotide 3-terminal de ladite molécule ADNc et facultativement en aval de ladite séquence ribozyme génomique il comprend au moins un terminateur, de préférence le terminateur T7.
 12. Procédé selon la revendication 11, dans lequel la séquence ribozyme génomique est la séquence ribozyme génomique du virus delta de l'hépatite.
 13. Procédé selon l'une quelconque des revendications 4 à 12, dans lequel ledit plasmide est capable de se répliquer dans un hôte eucaryote ou procaryote.
 14. Procédé selon l'une quelconque des revendications 5 à 13, dans lequel le fragment d'ADN exprimable dudit plasmide est un fragment d'ADN qui est homologue ou hétérologue par rapport au virus ARN à brin négatif et codant au moins un épitope immunogène.
 15. Procédé selon la revendication 14, dans lequel le fragment d'ADN exprimable dudit plasmide code au moins un épitope immunogène d'au moins un pathogène, de préférence une protéine enveloppe, ou au moins un produit génique ne possédant pas d'individus génétiquement déficients ou toxique pour des cellules malignes ciblées.
 16. Procédé selon la revendication 15, caractérisé en ce que ledit fragment d'ADN exprimable dudit plasmide est dérivé à partir d'un virus, d'une bactérie ou d'un parasite.
 17. Procédé selon l'une quelconque des revendications 5 à 16, dans lequel ledit fragment d'ADN exprimable dudit plasmide code un épitope immunogène capable d'obtenir une réponse immunoprotectrice.
 18. Procédé selon l'une quelconque des revendications 2 à 17, dans lequel le virus ARN à brin négatif est le virus de la rougeole ou des oreillons.
 19. Procédé selon l'une quelconque des revendications 1 à 17, dans lequel, dans la cellule auxiliaire lesdits gènes codant la protéine N, P et L sont dérivés du

virus de la rougeole et des oreillons.

20. Procédé selon l'une quelconque des revendications 1 à 19, dans lequel ladite cellule auxiliaire est dérivée de la lignée des cellules embryonnaires des reins humains 293 (ATCC CRL 1573). 5
21. Procédé selon l'une quelconque des revendications 1 à 20, dans lequel le rapport entre le plasmide tel que défini dans l'une quelconque des revendications 4 à 20 et le plasmide comprenant l'ADN codant la protéine virale L est d'environ 1000 : 1. 10
22. Procédé selon l'une quelconque des revendications 1 à 21, dans lequel ladite récupération dudit virus est réalisée directement à partir de la culture de cellules auxiliaires transfectées après formation syncytiale. 15
23. Procédé selon l'une quelconque des revendications 1 à 22, dans lequel ladite récupération dudit virus est obtenue après mélange de la cellule auxiliaire transfectée avec d'autres cellules aptes à être infectées et capables d'effectuer la réplication dudit virus. 20 25
24. Virus ARN à brin négatif non segmenté, infectieux de la famille Paramyxoviridae obtenu par le procédé selon l'une quelconque des revendications 7 à 10. 30
25. Vaccin comprenant le virus ARN selon la revendication 24, facultativement en combinaison avec un excipient pharmaceutiquement acceptable. 35

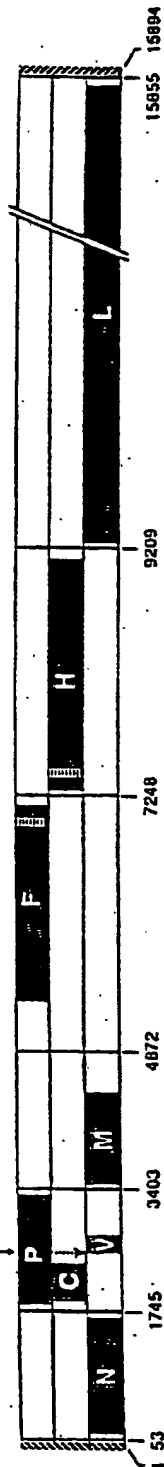
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
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G Insertion
2499



Symbols		Gene numbering (nt) (numbers in brackets: counted from transcription start)				
		Transcription		Translation		
		Initiation	Termination	Start (first nt of start codon)	Stop (last nt of stop codon)	
	= leader					
	= untranslated region (UTR)					
	= gene boundary trinucleotide					
	= open reading frame (ORF)					
	= change of ORF					
	= transmembrane coding region					
	= trailer					
	= position of the first residue of the gene boundary trinucleotide					
		N gene	56 (1)	1744 (1689)	108 (53)	1685 (1630)
		P gene	1748 (1)	3402 (1655)	1807 (60)	3330 (1583)
		C			1829 (82)	2389 (642)
		V			1807 (60)	2705 (950)
		M gene	3406 (1)	4871 (1466)	3438 (33)	4445 (1040)
		F gene	4875 (1)	7247 (2373)	5449 (575)	7110 (2236)
		H gene	7251 (1)	9208 (1958)	7271 (21)	9124 (1874)
		L gene	9212 (1)	15854 (6643)	9234 (23)	15785 (6574)

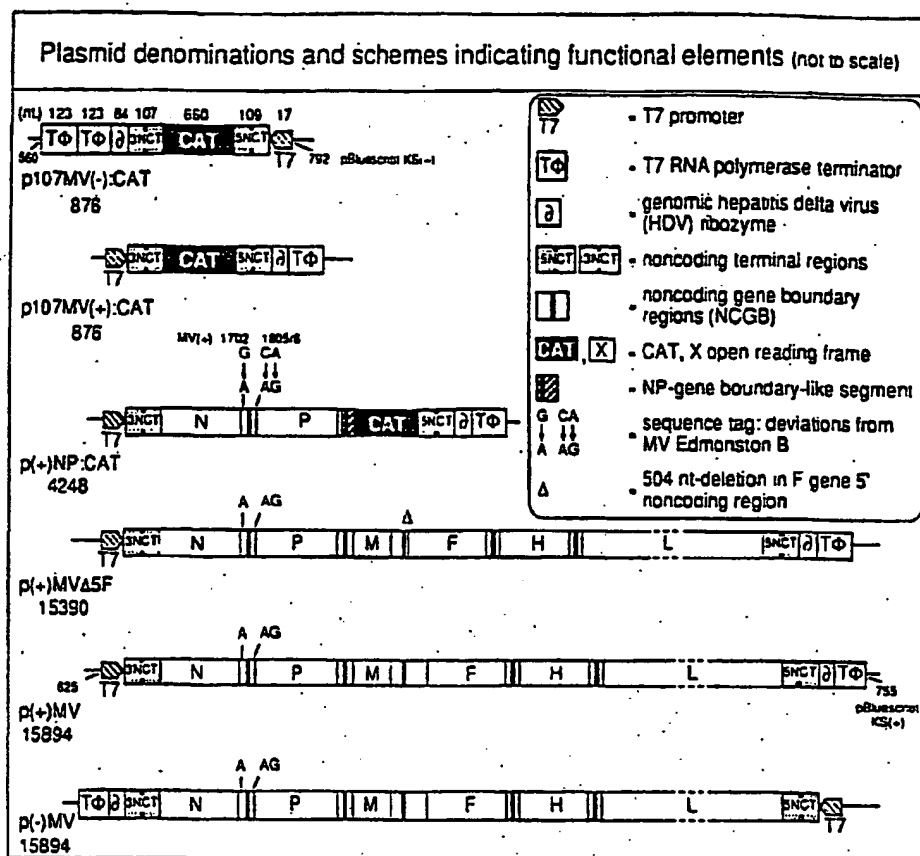


FIGURE 2

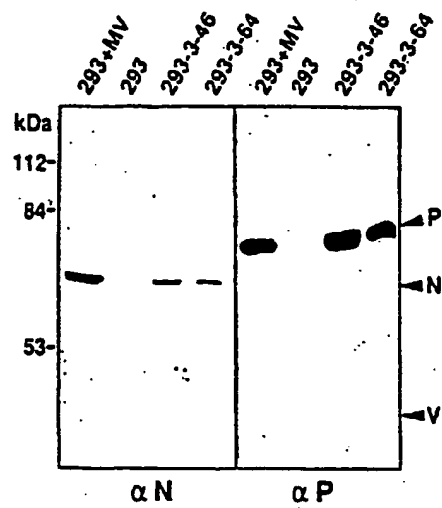


FIGURE 3

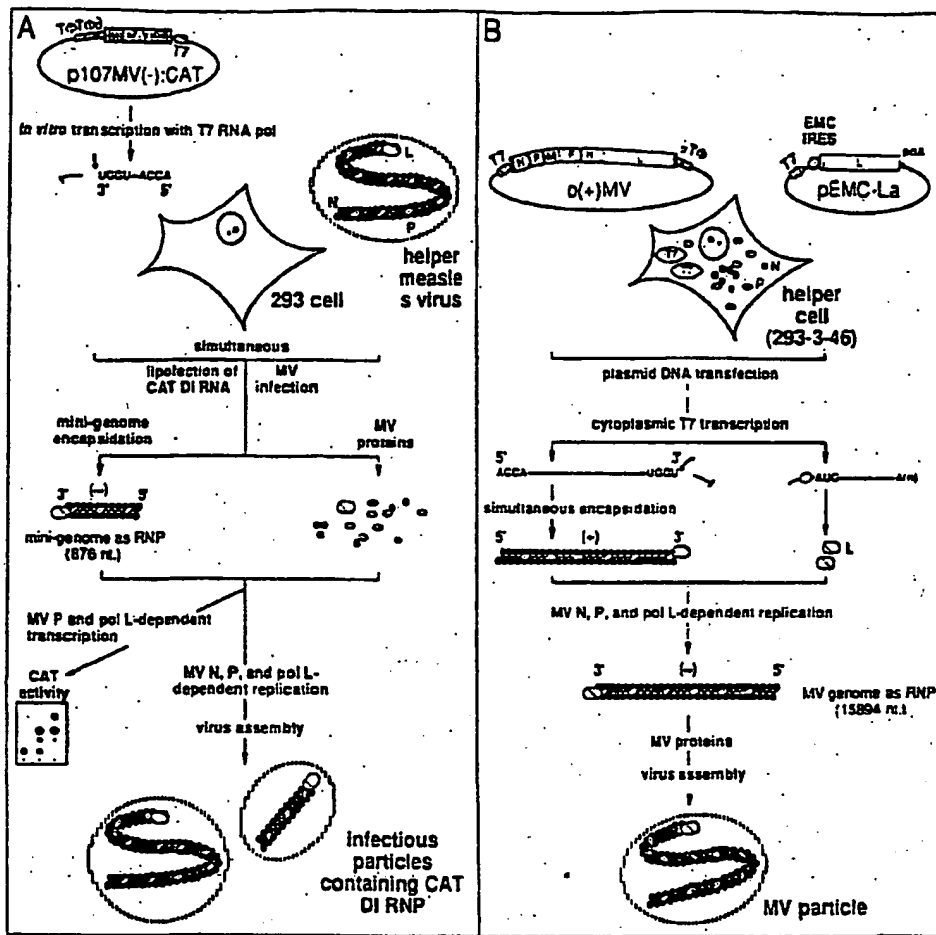


FIGURE 4

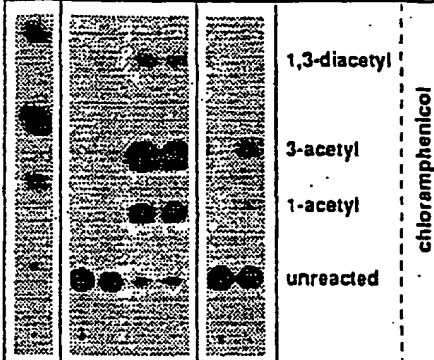
CAT control	ug of plasmid per 35 mm well					
	1	1	1	1	1	pT7P2lacZ
	5	5				p107MV(-):CAT
	5	5				p107MV(+):CAT
			5	5		p(+):NP:CAT
	0.1	0.1		0.1		pEMC-La
						
						1,3-diacetyl
						3-acetyl
						1-acetyl
						unreacted
						chromaphenicol

FIGURE 5

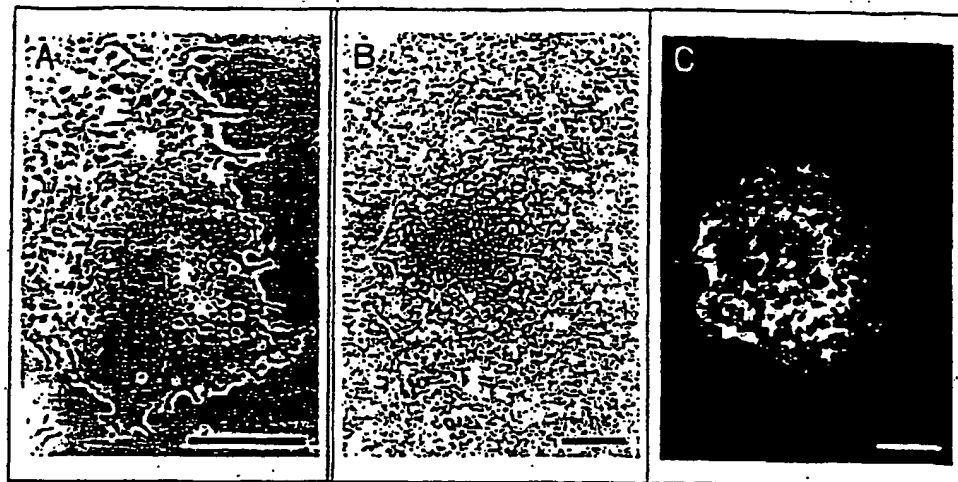


FIGURE 6

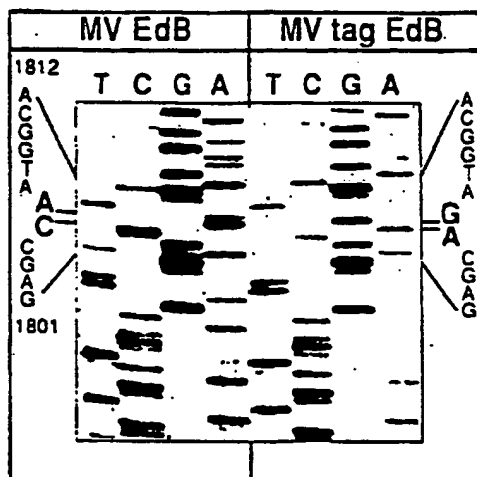


FIGURE 7

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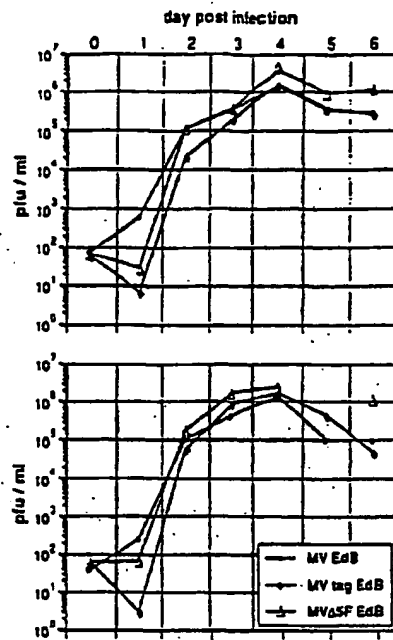


FIGURE 8

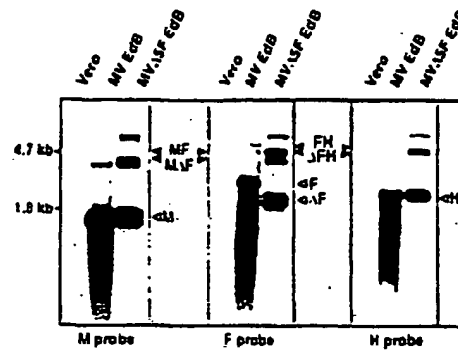


FIGURE 9

Plasmids for production of standard and deleted MVs and hybrid MVs containing additional genes or exchanged envelope proteins

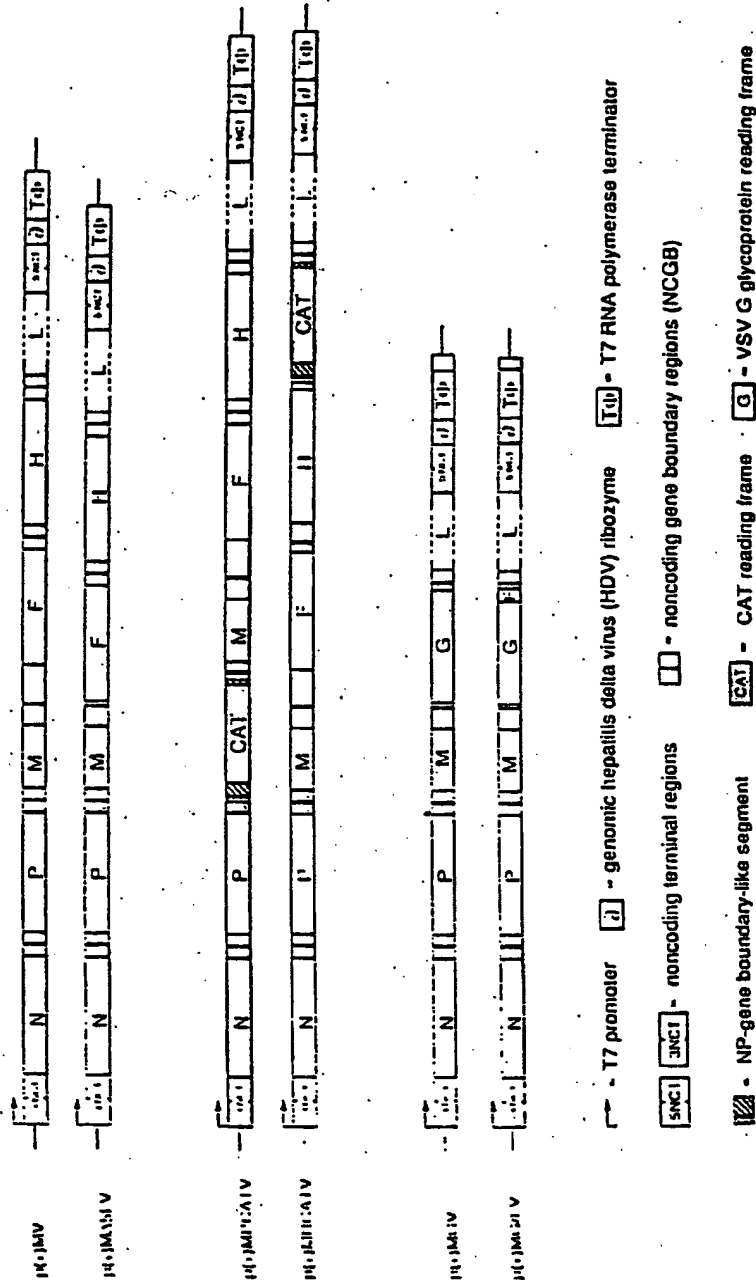


FIGURE 10

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Electron Microscopy of BHK cells infected with
replicating agent rescued from p(+)MGV

RNP structures (magnification: 41'700x)

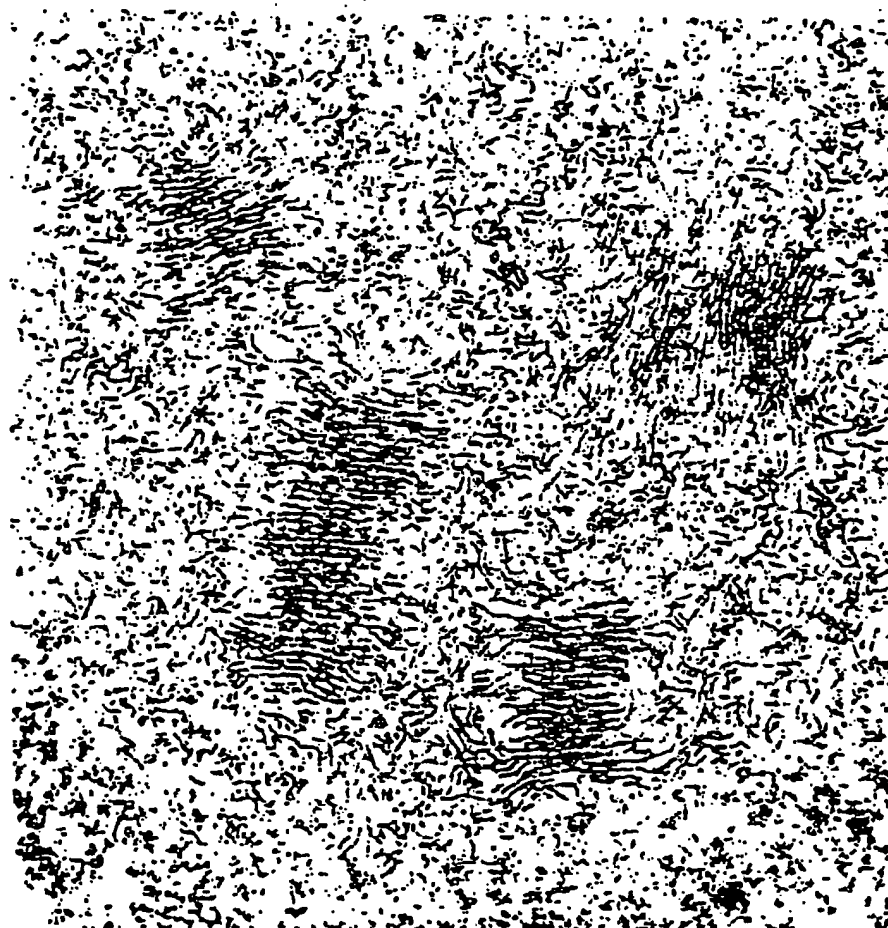


FIGURE 11

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Electron Microscopy of BHK cells infected with
replicating agent rescued from p(+)MGV

MV virion-like particles (magnification: 54'300x)



FIGURE 12

Electron Microscopy of BHK cells infected with VSV:

VSV virion particles (magnification: 41'700x)

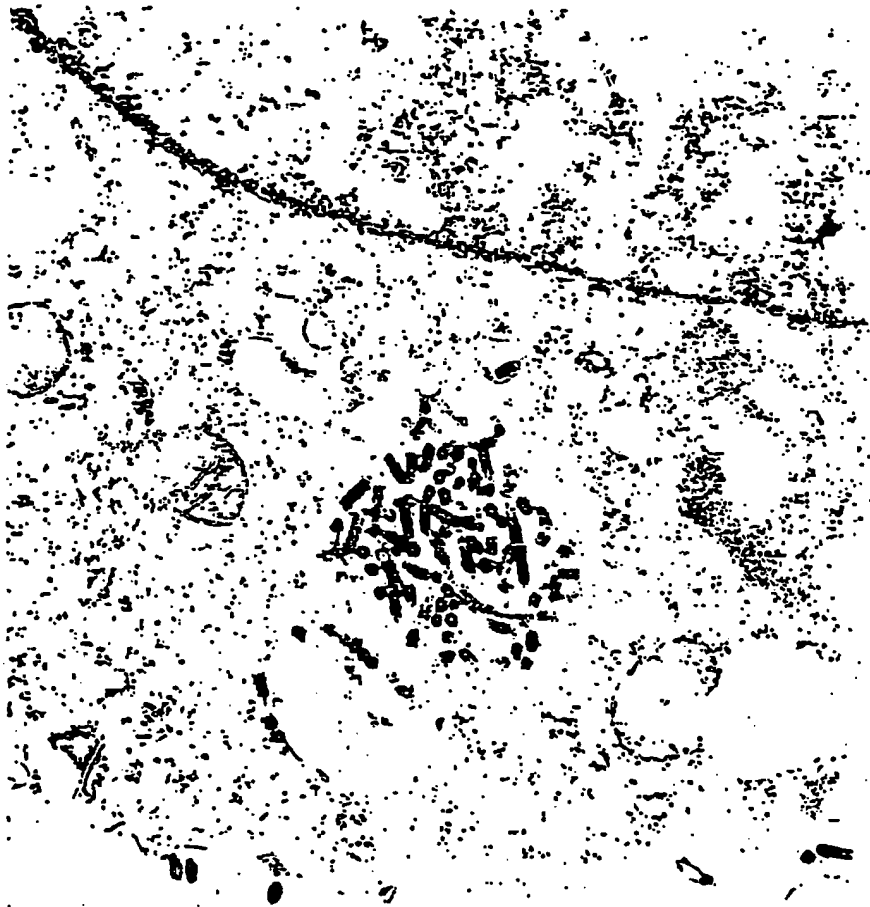


FIGURE 13

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